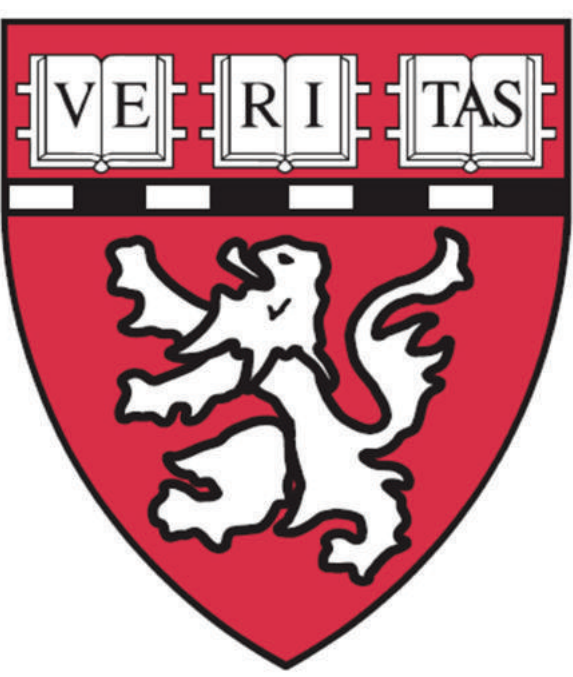




Aspergillus Melanin Potently Blocks Human Airway Epithelium Mediated Inflammation

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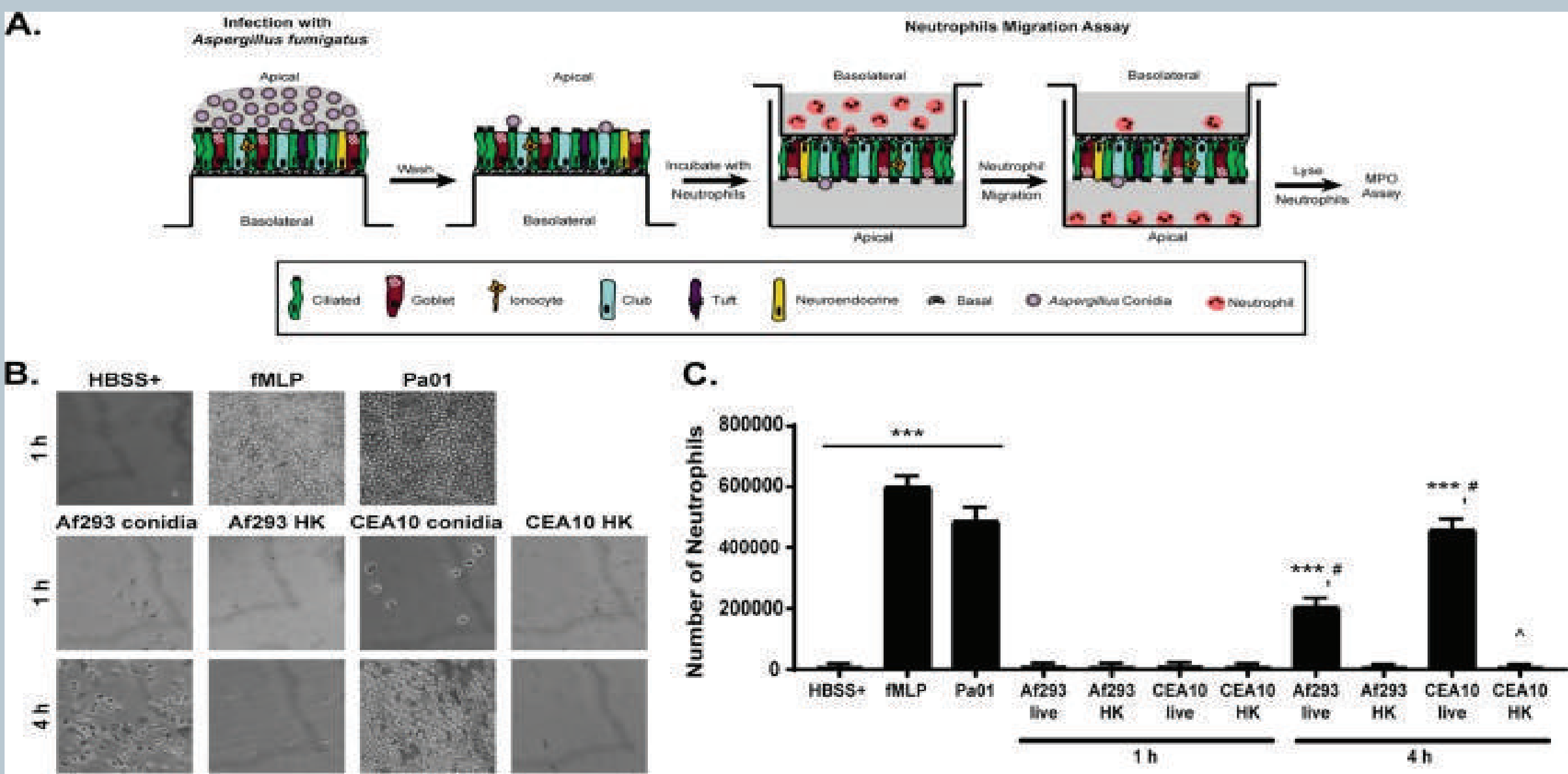
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ABSTRACT

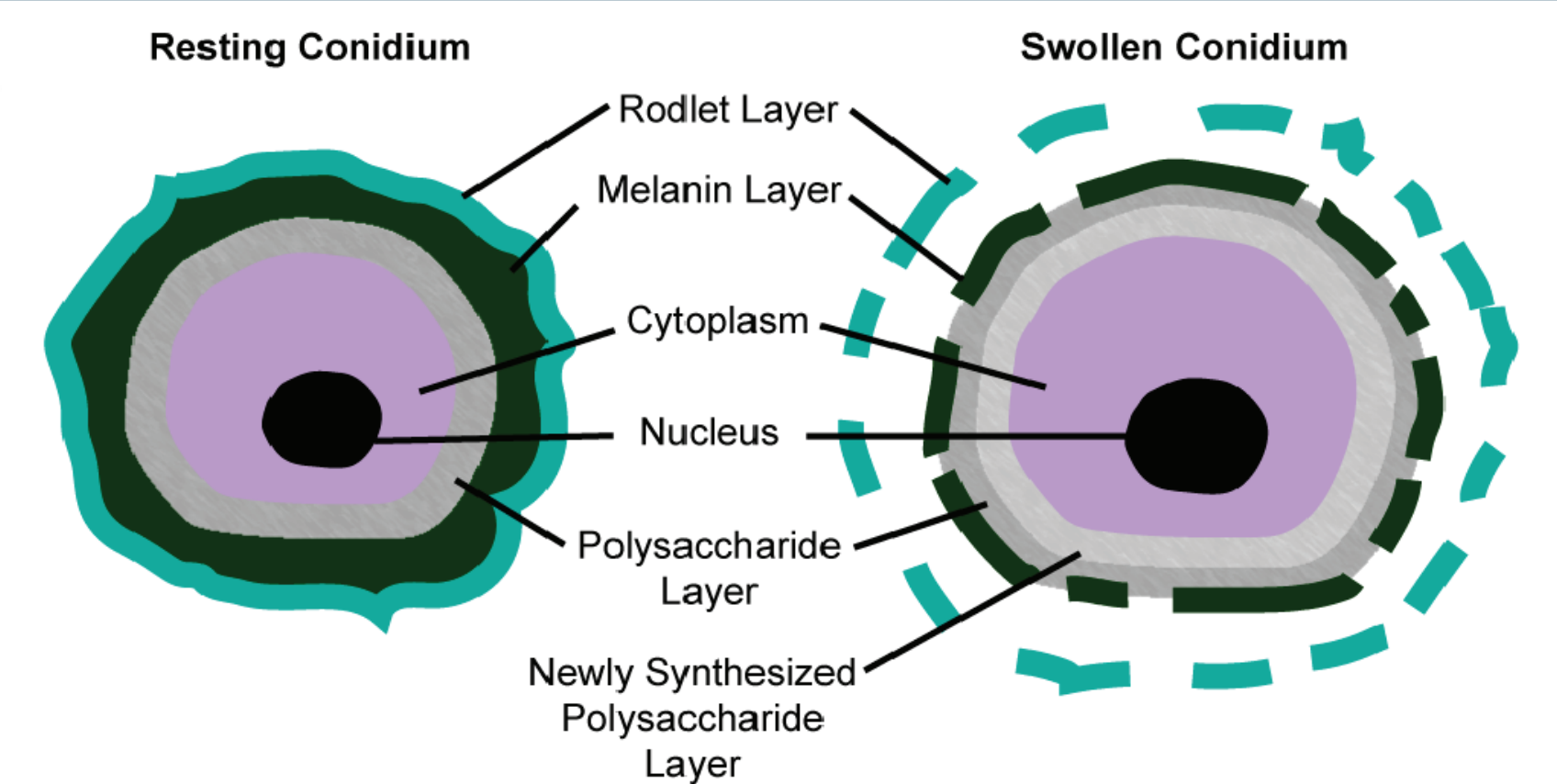
Most fungal pathogens, including *Aspergillus fumigatus*, are found in the environment and humans are exposed through the inhalation of fungal conidia. Conidia that remain in our lungs after exhalation are deposited onto the respiratory epithelium, particularly within terminal small airways. Clearance of fungal pathogens is multifactorial, relying upon mechanical means (clearance through the action of cilia and mucous) and through cell-mediated immunity, particularly via recruitment of innate immune cells to sites of infection. The airway epithelial is not just a passive barrier, but an active participant that is able to communicate with the immune system and trigger immune responses. However, the role of human airway epithelial cells (HAEC) in orchestrating protective or maladaptive immune responses is poorly understood. Initiation of a host-response depends upon recognition of the dynamic, multi-layered carbohydrate rich fungal cell wall. In addition to the typical carbohydrate components, most fungal conidia, including *Aspergillus fumigatus*, also contain a melanin layer that provides protection against environmental stresses, but it's role in modulating the airway drive immune response has not been explored. Using both a human muco-epithelial cell line, NCI-H292, and a fully pseudostratified primary Human airway epithelial cell model grown at air-liquid interface, we demonstrate that *A. fumigatus* melanin potently inhibits transmigration of neutrophils across the airway epithelium. *Aspergillus* melanin blocks the production of pro-inflammatory neutrophil chemoattractant, CXCL8 (IL-8) and CXCL1 (Gro-alpha) by selectively targeting apical secretion into the airways, resulting in a failure to generate a chemokine gradient that triggers neutrophil influx (or efflux) into the airways. Our results demonstrate that melanin actively down-regulates airway epithelial mediated pro-inflammatory responses toward *Aspergillus* and reveals a new strategy by which *aspergillus* subverts the immune system.

Prolonged Infection with live WT *Aspergillus fumigatus* is required for transepithelial migration of PMNs

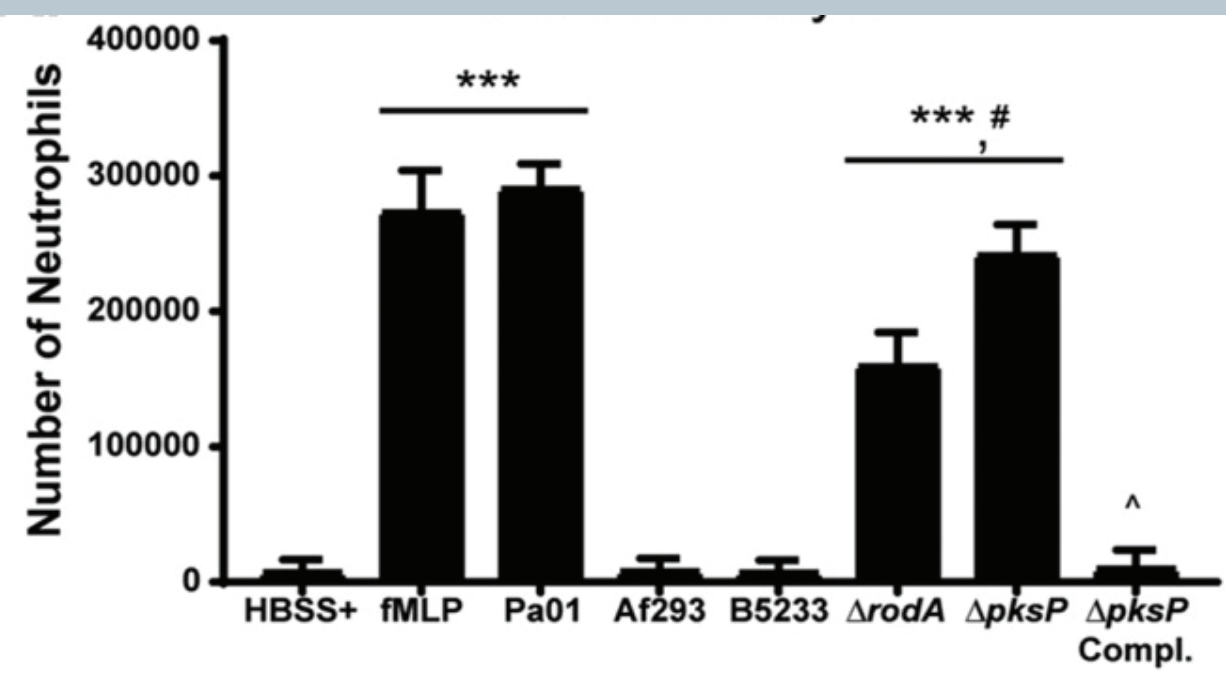


Prolonged epithelial exposure to live *A. fumigatus* promotes neutrophil migration. (A) Cartoon representation of an inverted ALI neutrophil transmigration experiment. (B and C) Light microscopy *B) and MPO assay quantification (C) of migrated neutrophils following epithelial stimulation with HBSS, 100nM fMLP, *P. aeruginosa* (PaO1), Af293 live and heat-killed (HK) resting conidia, and CEA10 live and HK resting conidia. All infections were performed with a 1×10^7 conidia/cm² of epithelial surface area for 1 hour or 4 hours, as indicated. Error bars represent the SD of the mean. P values were determined by one-way ANOVA with Tukey's post hoc test for multiple comparisons (n=6). ***, p < 0.0001 versus the results for HBSS control; #, p < 0.0001 versus the results at 1 hour; ^, p < 0.0001 versus the results for live conidia. (Feldman, et al. Infect Immun. 2020 Feb; 88(2): e00813-19).

Active Remodeling of the *Aspergillus* Cell Wall During Conidial Germination

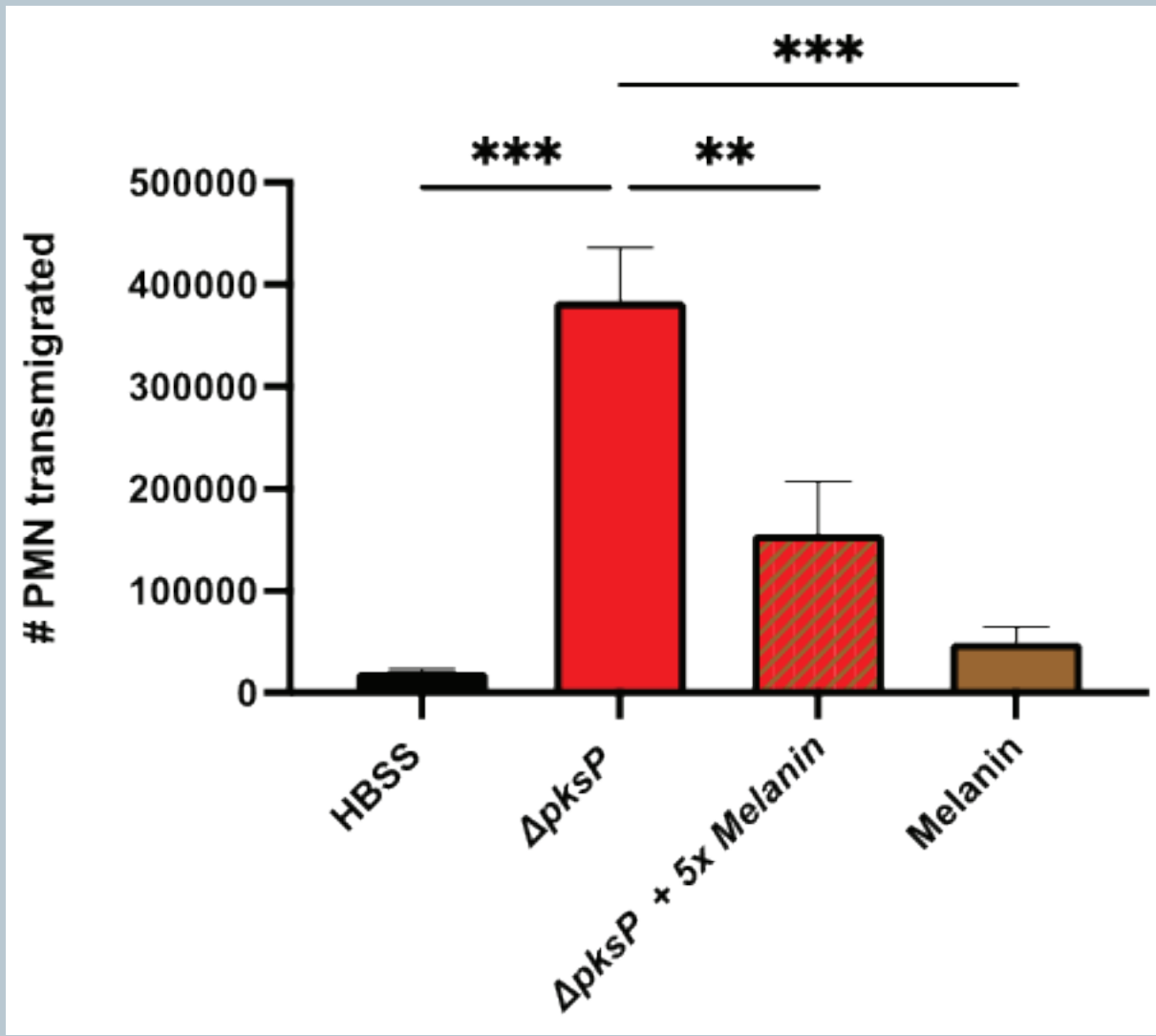


Melanin Deficient *Aspergillus* Promotes Early Robust PMN Transmigration



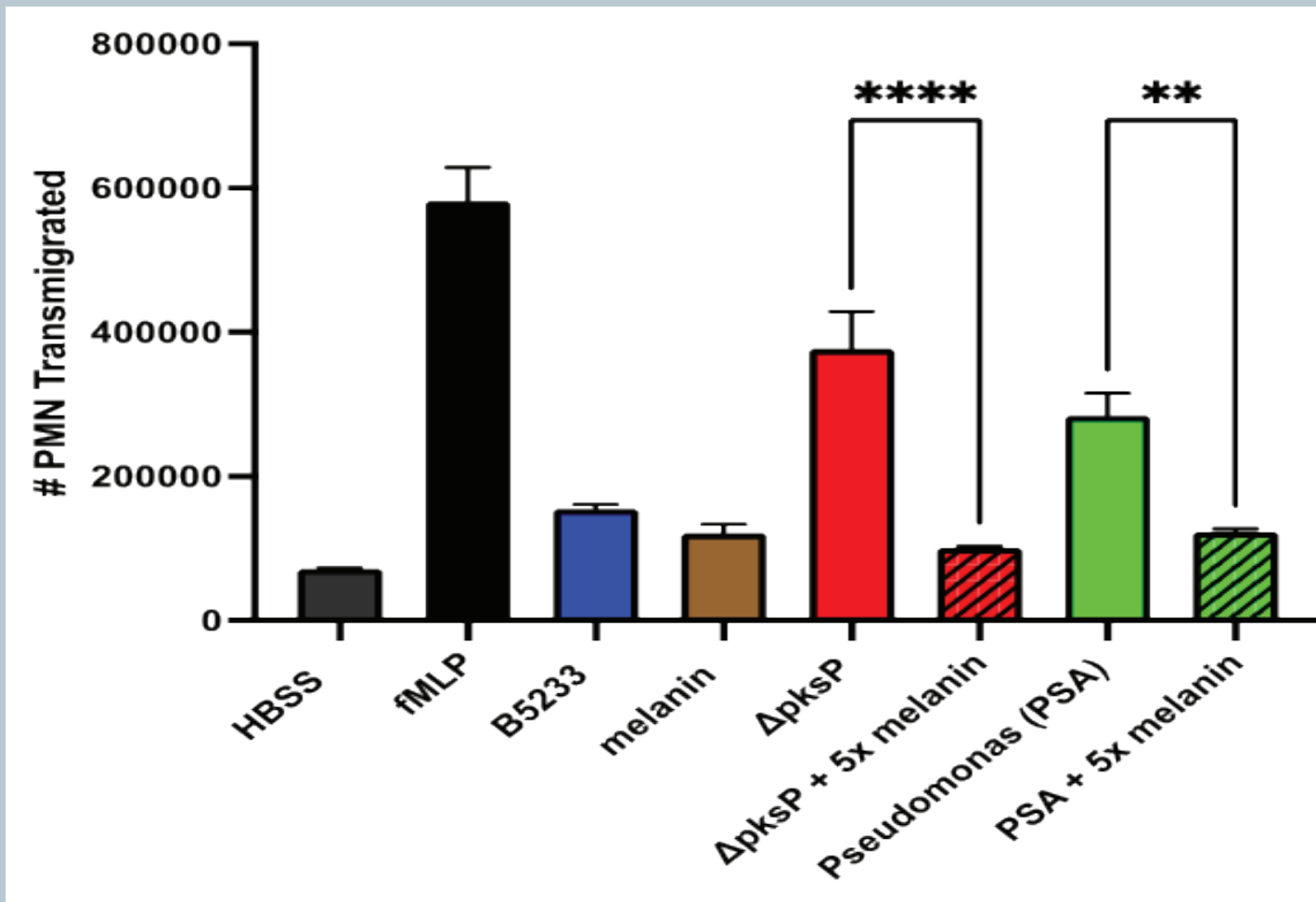
Conidia of *A. fumigatus* Δ rodA and Δ pksP stimulate rapid PMN migration across epithelium. PMN migration to the apical surface of H292 epithelium following stimulation with 1×10^7 conidia/cm² of *A. fumigatus* conidia from WT (B5233, Af293), Δ rodA, Δ pksP, and Δ pksP-complemented strains for 1 hour. Error bars represent the SD of the mean. P values were determined using one-way ANOVA with Tukey's post hoc test for multiple comparisons. ***, p < 0.0001 versus the results for HBSS control; #, p < 0.0001 versus the results for the WT; ^, p < 0.0001 versus the results for the Δ pksP mutant conidia (Feldman, et al. Infect Immun. 2020 Feb; 88(2): e00813-19).

A. fumigatus Melanin Ghosts Suppress Transepithelial PMN Migration



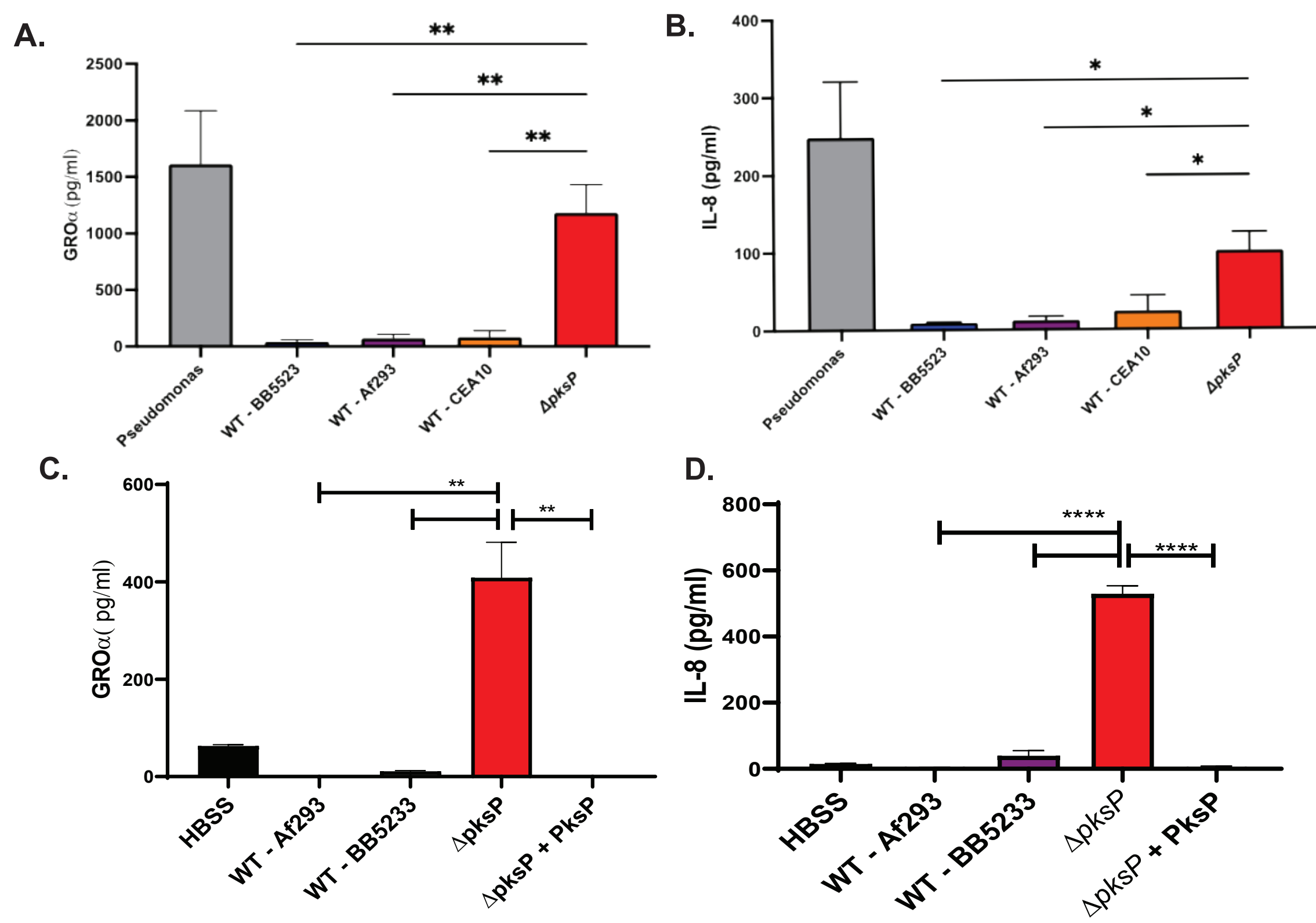
Primary HAEC derived from healthy donor basal cells was plated on 24-well transwells and allowed to differentiate for 16 days into a fully pseudostratified epithelium at air-liquid interface (ALI) Melanin ghosts were created from wildtype strain B5233. Epithelium were stimulated with HBSS, 1×10^7 conidia/cm² from Δ pksP strain or 1×10^7 Δ pksP conidia/cm² with 5×10^7 melanin ghosts/cm² or 5×10^7 melanin ghosts/cm² alone for 1 hour. PMN transmigration was measured using a myeloperoxidase assay. P values were determined by one-way ANOVA with Tukey's post hoc test for multiple comparisons; ***, p < 0.0001, **, p < 0.001.

Melanin Suppresses PMN Transepithelial Migration in Response to both Fungal and Bacterial Pathogens



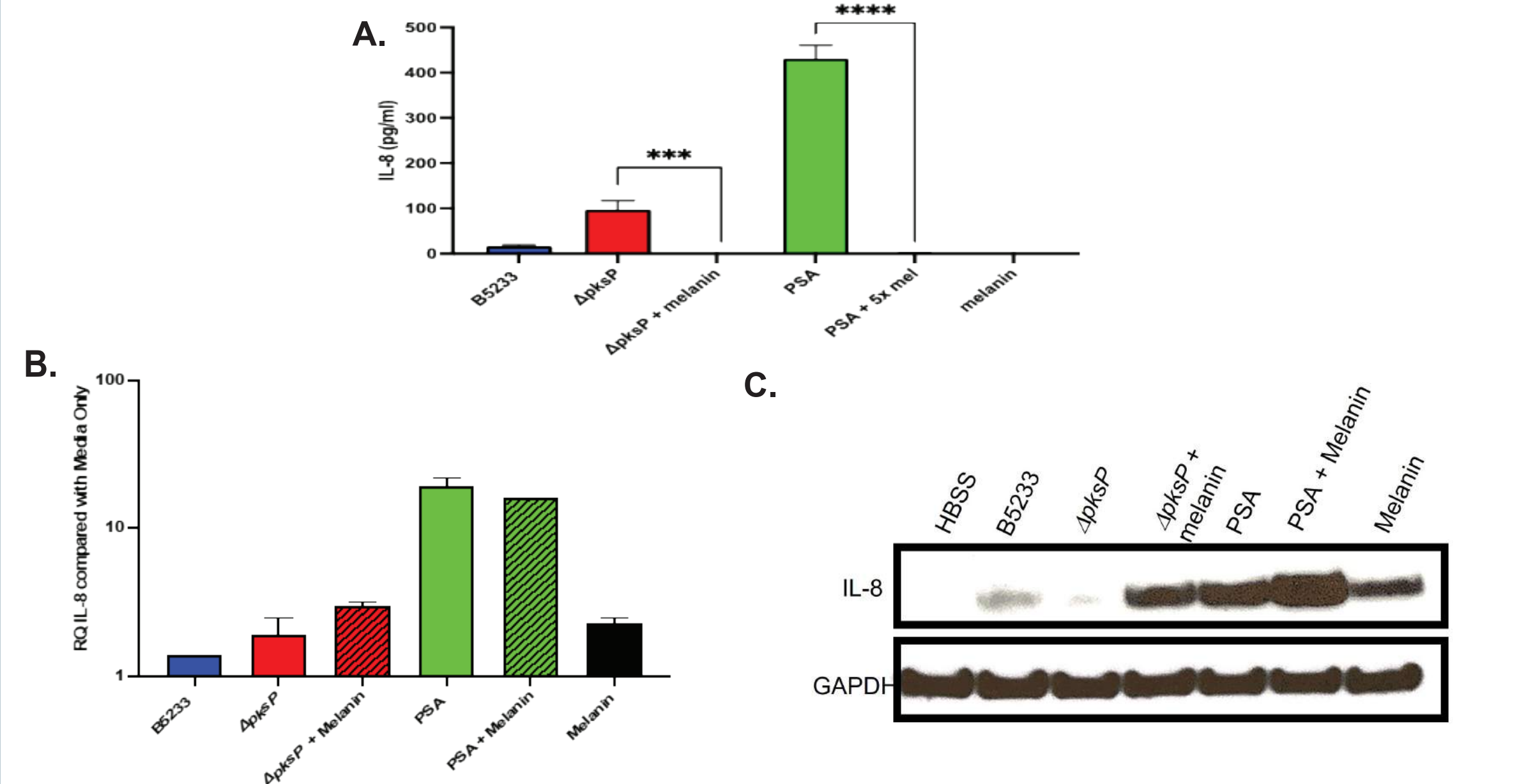
HAEC were stimulated with *Aspergillus* wildtype (B5233), Δ pksP conidia at 1×10^7 conidia/cm² or *P. aeruginosa* (PaO1) alone or in combination with 5×10^7 melanin ghosts/cm² in HBSS media. fMLP was used as a positive control for PMN transmigration. Transmigration was quantified using a myeloperoxidase assay and compared with a standard curve. Error bars represent standard error of three biologic replicates, statistics were calculated with two-way ANOVA using PRISM 9 software (** p $\leq$ 0.01; **** p < 0.0001).

A. fumigatus Δ pksP induces PMN Chemoattractants IL-8 and Gro- α in both HAEC and NCI-H292 epithelium



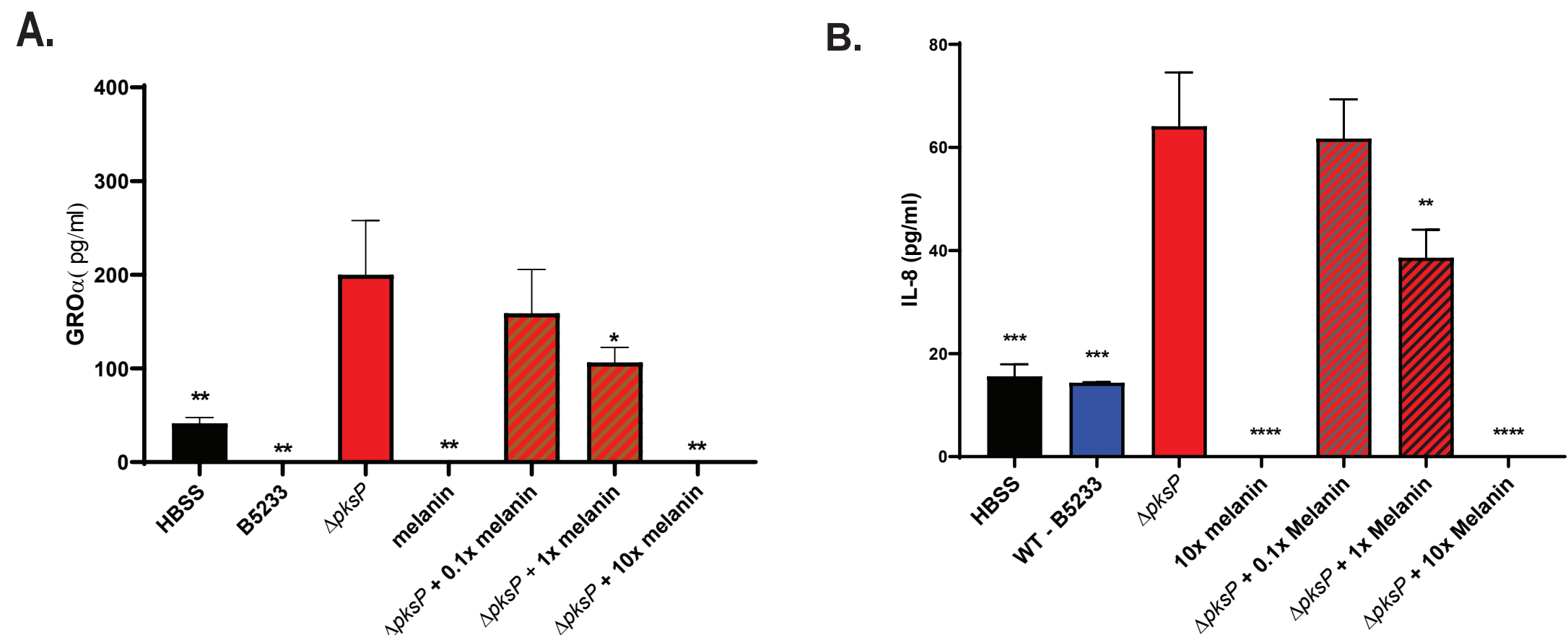
(A and B) Primary HAEC derived from three healthy were stimulated with *A. fumigatus* wildtype strains (B5233, Af293, CEA10), Δ pksP conidia or *P. aeruginosa* strain PaO1. Fungal stimulation were performed at 1×10^7 conidia/cm² for 4 hours and cytokines were measured using Luminex. (C and D) Confluent epithelia composed of NCI-H292 cells were stimulated using the same conditions as the HAEC and cytokine production was measured using ELISA. Error bars represent the standard deviation of the mean. P values were determined by one-way ANOVA with Tukey's post hoc test for multiple comparisons; ****, p < 0.00001; ***, p < 0.0001, **, p < 0.001, *, p < 0.01.

Aspergillus Melanin Blocks Secretion, But Not Translation or Transcription of IL-8



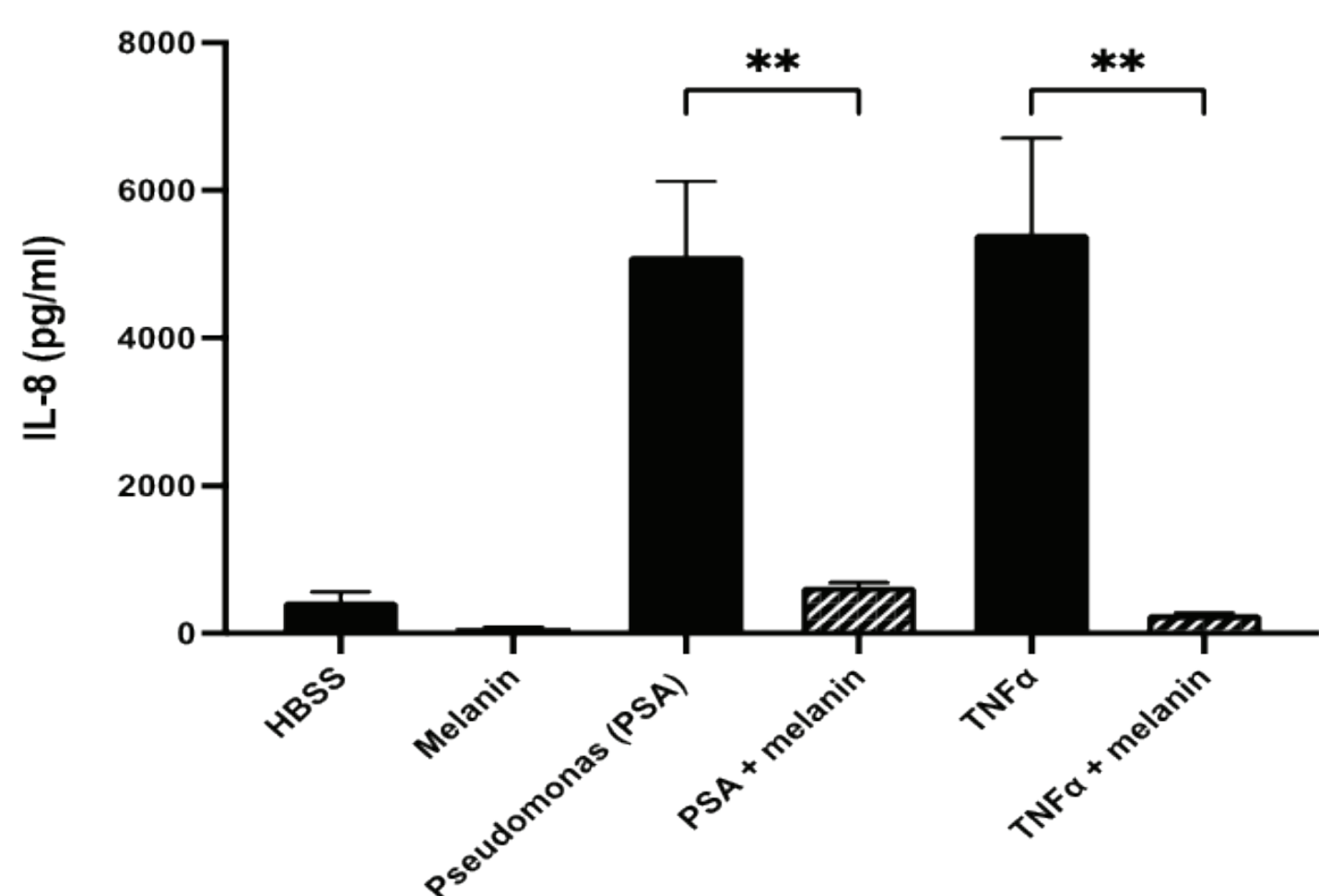
Confluent NCI-H292 monolayers grown on transwells were stimulated with *Aspergillus* wildtype, Δ pksP, *P. aeruginosa* (PSA) at 1×10^7 conidia/cm² or in combination with melanin ghosts at 5×10^7 ghosts/cm². Supernatants from the apical compartment were analyzed for IL-8 using ELISA (R&D, Duoset). Five replicates were performed for each condition, three replicates were processed to purify total RNA and two replicates were lysed using 1% NP40 lysis buffer and processed for immunoblotting. (A) Quantification of IL-8 in the apical supernatant (B) Total RNA samples were converted to cDNA using Superscript IV Vilo MasterMix (ThermoFisher) and qPCR was performed using TaqMan probes for IL-8 and GAPDH (control) and the TaqMan Advanced Master Mix (ThermoFisher) and quantified using an Applied Biosystems 7500 Fast Real-Time PCR System. RQ was computed using manufacturers software and error bars represent the standard deviation from 3 biological replicates. (C) Immunoblotting of lysates for IL-8 and GAPDH.

Aspergillus Melanin Suppression of IL-8 and Gro- α is dose-dependent



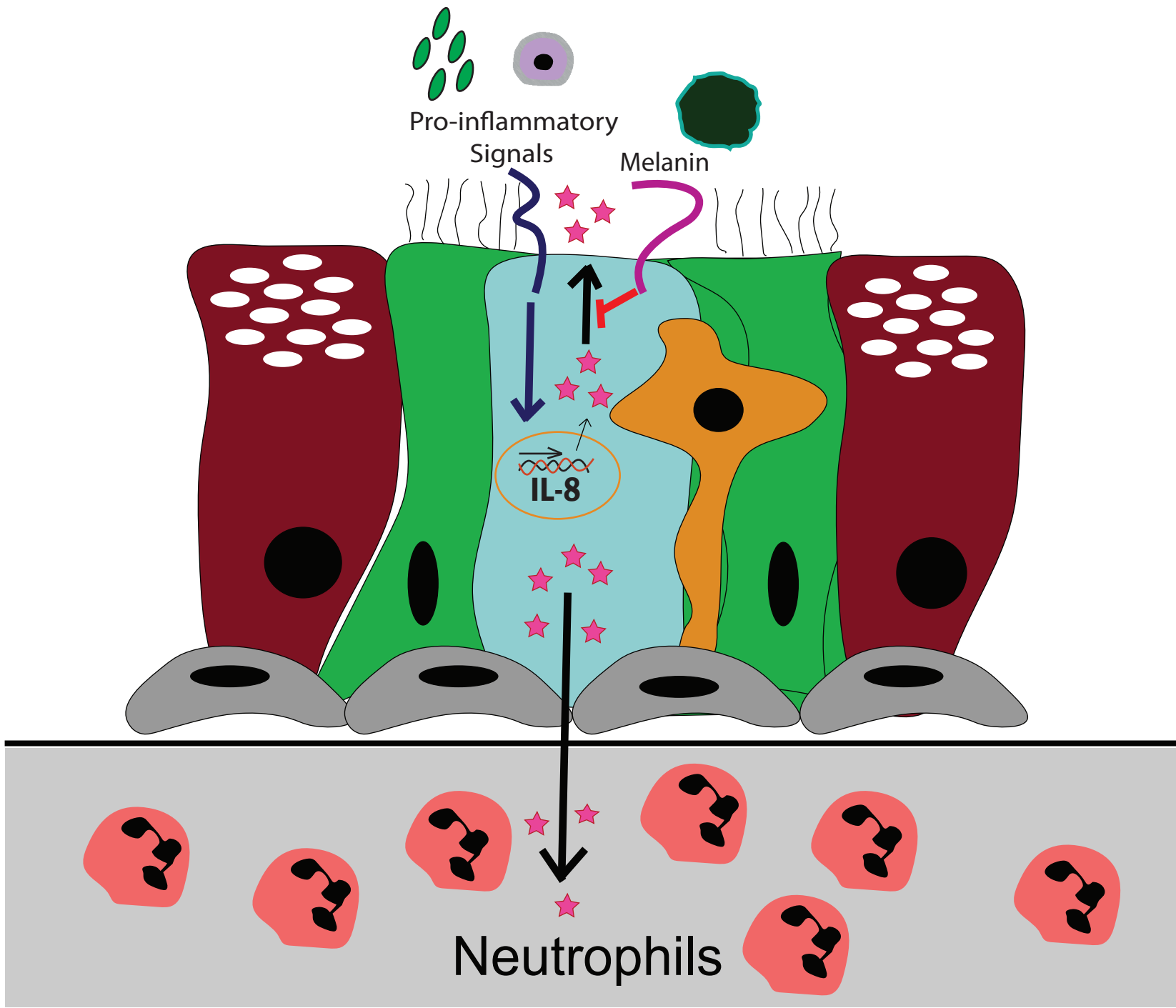
Primary HAEC was stimulated with *Aspergillus fumigatus* conidia from wildtype (B5233) or Δ pksP strains at 1×10^7 conidia/cm² and were mixed with melanin ghosts at the indicated concentration (relative to the Δ pksP strain). Supernatants were analyzed for Gro- α and IL-8 by ELISA (R&D, Duoset). Error bars represent SEM of three biological replicates, significance was determined by two-way ANOVA using PRISM 9 software. Significance indicated is relative to the Δ pksP strain (not significant, NS; *** p $\leq$ 0.001; **** p < 0.0001).

Aspergillus Melanin Suppresses Fungal, Bacterial and Cytokine Induced IL-8 Production



Primary HAEC was stimulated with wildtype *Aspergillus* melanin ghosts at 5×10^7 ghosts/cm², *P. aeruginosa* (PaO1), or 100 nM TNF α alone or in combination as indicated for 4 hours. Supernatants were harvested and analyzed for IL-8 using ELISA (R&D, Duoset). Error bars represent SEM of three biological replicates, significance was determined by two-way ANOVA using PRISM 9 software. (** p $\leq$ 0.01).

Conclusions



A. fumigatus wildtype strains induce delayed transepithelial migration of neutrophils, however strains that lack melanin (Δ pksP) promote early robust neutrophil transmigration.

Aspergillus melanin suppresses pro-inflammatory cytokine production by airway epithelial cells in a dose-dependent manner.

Melanin blocks both fungal, bacterial, and cytokine induced IL-8 production, and blocks both fungal and bacterial induced transepithelial migration of neutrophils.

Melanin does not block either transcription or translation of IL-8, but inhibits secretion of IL-8, revealing a novel role for melanin for dampening HAEC-derived inflammation in response to *Aspergillus*.