



EORTC / EFISG: Invasive aspergillosis management in 2017 and beyond

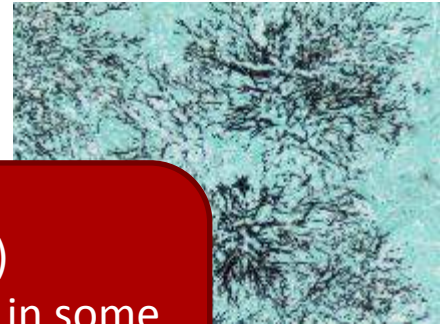
News from the lab

Ana Alastruey Izquierdo
Mycology Reference Lab Spain
Instituto de Salud Carlos III

Diagnosis of IA

Early diagnosis challenging and basic for improvement of survival

Gold standard  culture/direct sample/histopathology

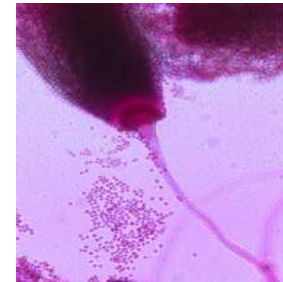
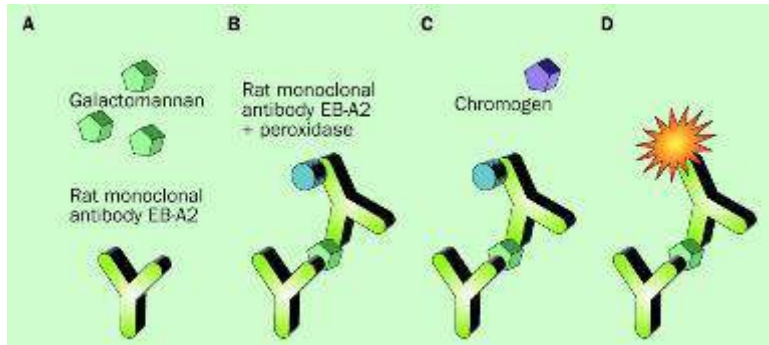


Biopsies difficult (critically ill)
Cultures low sensitivity and low PPV in some
populations
Antifungal treatment

Galactomannan



- ✓ Enzyme-linked immunoassay
- ✓ Serum BAL, CSF
- ✓ Can be used for monitoring the infection



JOURNAL OF CLINICAL MICROBIOLOGY, Oct. 1999, p. 3223-3228
0095-1137/99/\$04.00+0
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Vol. 37, No. 10

Autopsy-Controlled Prospective Evaluation of Serial Screening for Circulating Galactomannan by a Sandwich Enzyme-Linked Immunosorbent Assay for Hematological Patients at Risk for Invasive Aspergillosis

JOHAN MAERTENS,¹ JAN VERHAEGEN,² HILDE DEMUYNCK,¹ PENELOPE BROCK,³
GREGOR VERHOEF,¹ PETER VANDENBERGHE,¹ JOHAN VAN ELDERE,²
LUDO VERBIST,² AND MARC BOOGAERTS^{1*}

Departments of Haematology,¹ Paediatrics,² and Microbiology,³ University Hospital Gasthuisberg, Leuven, Belgium

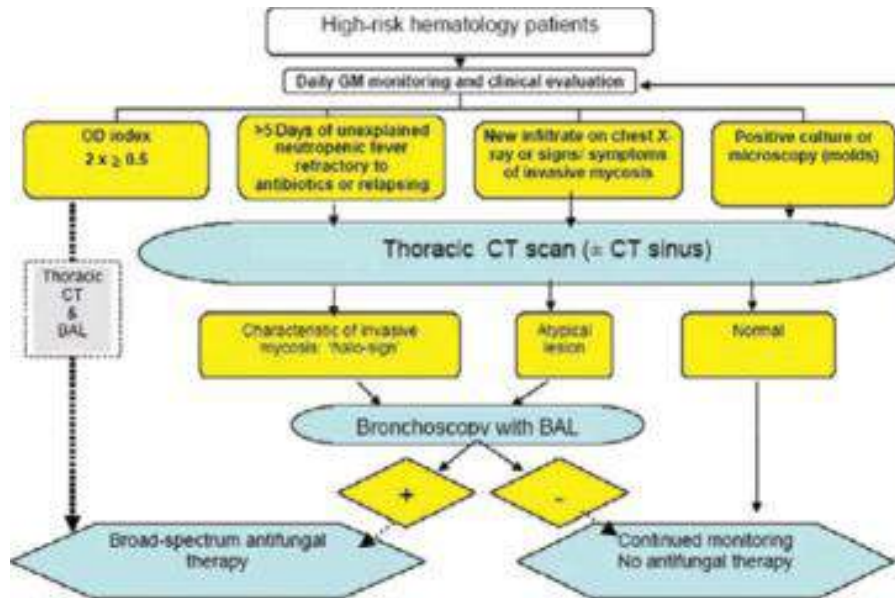
Bronchoalveolar Lavage Fluid Galactomannan for the Diagnosis of Invasive Pulmonary Aspergillosis in Patients with Hematologic Diseases

Johan Maertens,¹ Vincent Maertens,¹ Koen Theunissen,² Wouter Meersseman,² Philippe Meersseman,² Stet Meers,¹ Eric Verbeke,² Gregor Verhoef,¹ Johan Van Eldere,² and Katrien Lagrou¹

Sensitivity: 92.3%
Specificity: 95.4%

| Method | Sensitivity |
|------------|-------------|
| GM-BAL | 91.3% |
| Culture | 50% |
| Microscopy | 53.3% |

Galactomanan



Maertens J et al. Clin Infect Dis. 2005;41(9):1242-1250.

Reduction in empiric treatment 35 → 7,7%

- Other populations GM not performed well
 - ICU
 - COPD
 - AIDS
- Prophylaxis

| Population | Sensitivity | Specificity |
|------------------|-------------|-------------|
| Neutropenic | 96% | 96% |
| HCT | 70% | 91% |
| Liver transplant | 57% | 94% |
| ICU | 38% | 100% |
| CGD | 24% | 95% |

Adapted from Aguado GEMICOMED 2017

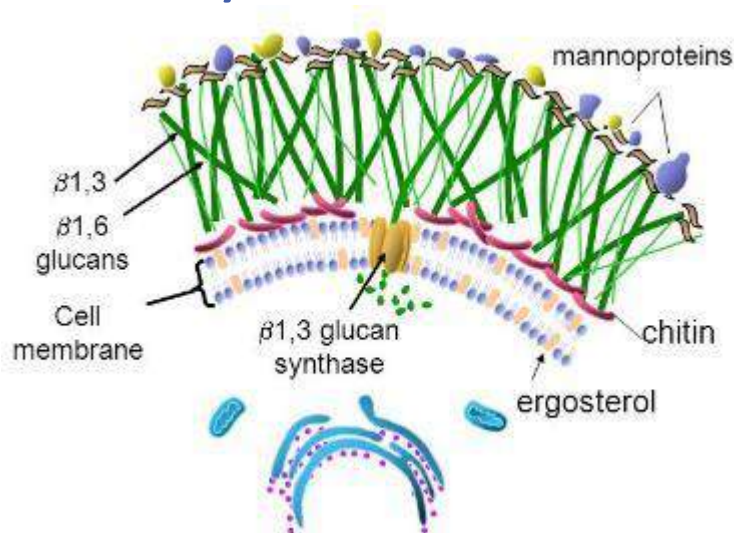
Maertens, Blood 01; Bretagne CID 98; Fortun, Transplant 01; Walsh, ICAAC 02

Table 2 Biological and epidemiological factors that influence the performance of GM detection in invasive aspergillosis³

| <i>Biological factors</i> | <i>Epidemiological factors</i> |
|--|---|
| Site of infection | Patient population |
| <i>Aspergillus</i> species causing infection | Sampling strategy |
| Microenvironment at the site of infection: nutrients, oxygen level, pH | Definition of a positive result |
| Exposure to antifungal agents | Definition of an IFD |
| Molecular structure of released galactomannan | Prevalence of IFD |
| Underlying condition/neutropenia/level of immunosuppression | Cutoff for positivity |
| Renal clearance, hepatic metabolism | Laboratory experience |
| Circulating galactomannan antibodies | Nutritional factors (galactomannan-containing food) |
| Storage of clinical sample | Treatment with semi-synthetic β -lactam antibiotics |
| Pre-analytical treatment procedure | |

β -D glucan

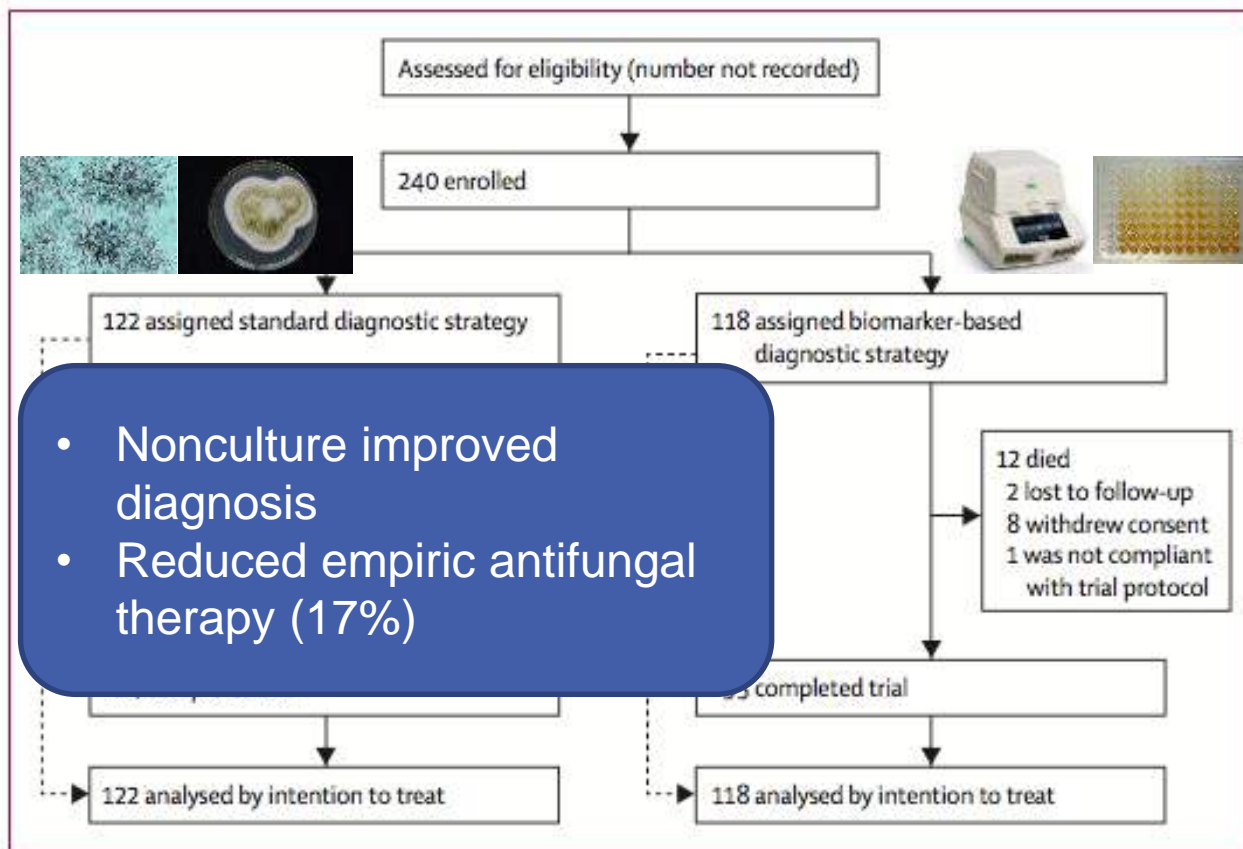
- “pan fungal” (No Crypto no Mucorales)
- Sensitive
- Low specificity



- Standardized
- Validation
- Variability
- EAPCRI --> DNA extraction

Biomarkers vs Standard

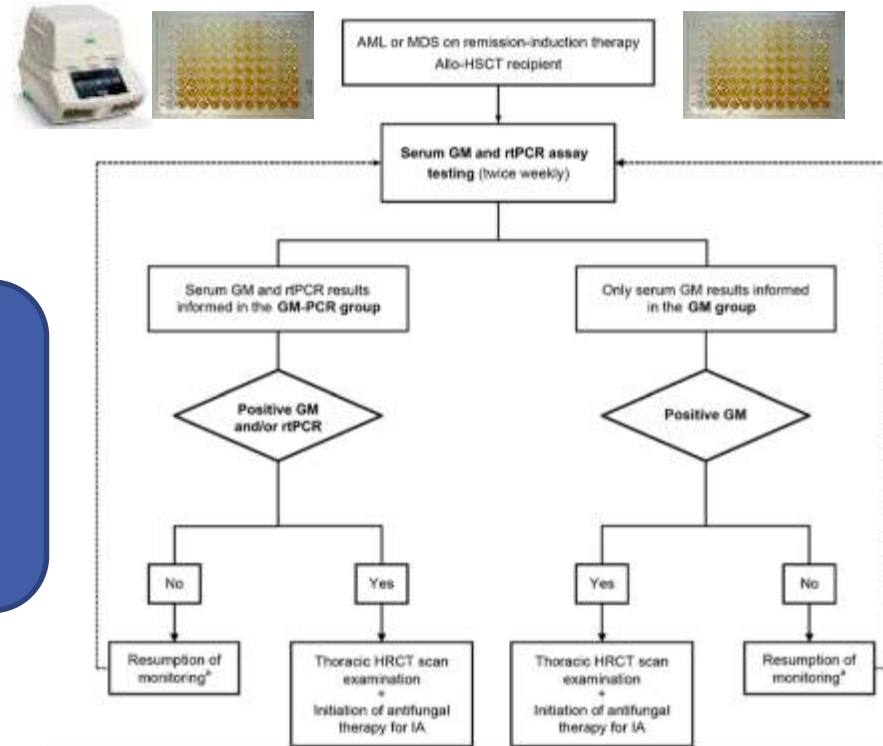
Multicenter
randomized
controlled
trial



GM vs GM+PCR

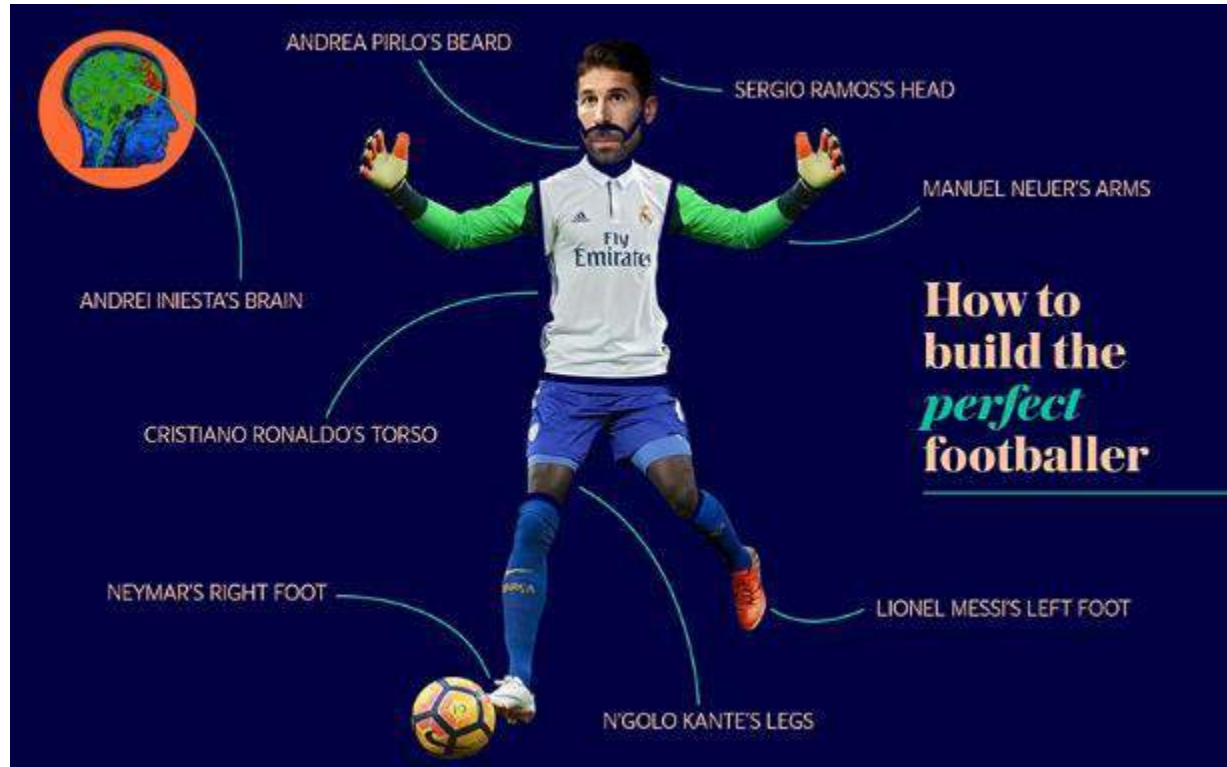
Multi- center randomized study
PCRAga

- ✓ Median to diagnosis GM-PCR (13 vs 20 days $P= 0.022$)
- ✓ Reduced empiric antifungal therapy (17% vs 29% $P= 0.027$)



No one is perfect

□ Combinations



No one is perfect

- Combinations
- New methods needed but...
- No reliable gold standard for validation
- EORTC/MSG
 - Proven: culture or histology ster
 - Probable: host + clinical + microbi
 - Possible: host + (clinical or microbiological)



Miss classification possible
Source of variability

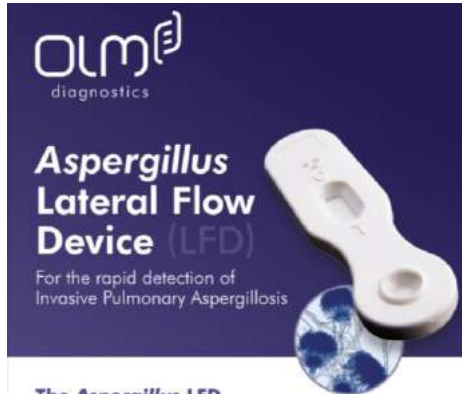


Lateral Flow assay



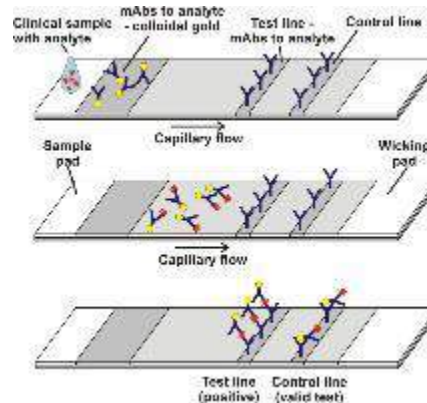
Alastruey-Izquierdo TIMM 2017

- Monoclonal Ab
- Detects extracellular glycoprotein
- Secreted during active growth of *Aspergillus* spp



The *Aspergillus* LFD

- Highly specific and detects 'activity' only
- Single use assay with results in less than 15 minutes.
- Proven efficacy in diagnosis of IPA in humans (serum and BAL)



PERFORMANCE CHARACTERISTICS

1. In haematological malignancy patients using serum EORTC Proven/probable IPA (n = 22) versus no IFD (n = 59)

Published study conducted at School of Medicine, Cardiff University, Wales, UK6. LFD compared to Bio-Rad Platelia GM-EIA and real-time PCR.

| ASSAY | SENSITIVITY | SPECIFICITY | NPV |
|--------|---------------------|---------------------|---------------------|
| LFD | 81.82% (61.5-92.7)* | 84.75% (73.5-91.8)* | 92.59% (82.5-97.1)* |
| PCR | 95.45% (78.2-99.2)* | 72.88% (60.4-82.6)* | 97.73% (88.2-99.6)* |
| GM-EIA | 77.27% (56.6-89.9)* | 81.36% (69.6-89.3)* | 90.60% (79.8-95.9)* |

* = 95% Confidence Intervals, NPV = Negative Predictive Value

2. In haematological malignancy patients and solid organ transplant recipients using BAL (n = 37; 27 HM, 10 SOT; EORTC probable IPA n = 12)

Published study conducted at Medical University of Graz, Austria4. LFD compared to Bio-Rad Platelia GM-EIA.

| SENSITIVITY | SPECIFICITY | NPV |
|-------------|-------------|------|
| 100% | 81% | 100% |

NPV = Negative Predictive Value

GM levels in LFD negative BALs were significantly lower than in LFD positive BALs (n = 22; median <0.4 ODI; [IQR] <0.4-<0.4 vs. n = 17; median 1.50 ODI; [IQR] 0.72-1.13; p < 0.0001, Mann-Whitney U test). GM levels were also significantly lower in samples with weak LFD positives (n = 8; median 0.97 ODI; [IQR] <0.4-1.23), than in moderate or strong positive LFD samples (n = 9; median 4.66 ODI; [IQR] 2.8-19.3; p = 0.0012, Mann-Whitney U test).

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1. Thornton CR (2013). Lateral-flow device for diagnosis of infection. *Current Fungal Infection Reports*, published online. DOI: 10.1007/s12281-013-0136-4
2. Thornton CR (2008). Development of an immunochromatographic Lateral-Flow Device for Rapid Serodiagnosis of Invasive Aspergillosis. *Clinical and Vaccine Immunology* 15: 1099-1100.
3. Thornton CR, Johnson G, Agrawal S (2012). Detection of invasive pulmonary aspergillosis in haematological malignancy patients by using lateral-flow technology. *Journal of Hospital Infection* 81: e2923.
4. Hwang YM, Kahl C, Dwellmann W, Seiler K, Wagner J, Buzari W, Weller A, Rappert RB, Thornton CR (2012). Bronchoalveolar lavage lateral-flow device test for invasive aspergillosis diagnosis in haematological malignancy and solid organ transplant patients. *Journal of Infection* 65: 888-891.
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6. Hill J, Schweb T, Thornton CR, Koster E, Sans H (2013). Comparison of a Novel Aspergillus Lateral-Flow Device and the Platelia® Galactosyltransferase Assay for Diagnosis of Invasive Aspergillosis following Haematological Stem Cell Transplantation. *Infection*, published online. DOI: 10.1007/s001401304750

Lateral Flow assay



Alastruey-Izquierdo TIMM 2017

- ✓ Serum (pretreatment) and BAL samples
- ✓ Fast and cheap
- ✓ Sensitivity 68% (52-81%) specificity 87% (80-92)
- ✓ Cross-reaction with *Penicillium*
- ✓ Fast (15 min) and cheap
- ✓ Lower sensitivity in prophylaxis and patients with hematological malignancies

Table 3 Sensitivity of BALF LFD for probable/proven IPA in patients with and without antifungal prophylaxis/therapy (information only available for a proportion of cases published).^a

| | BALF LFD sensitivity for IPA overall percentage (absolute numbers) |
|--|--|
| Overall | 75% (50/67) |
| Under mold active systemic antifungals | 56% (14/25) |
| Without mold active antifungals | 86% (36/42) |

Table 1 Per BALF sample performance of the BALF *Aspergillus* LFD for probable/proven invasive pulmonary aspergillosis versus no evidence for invasive pulmonary aspergillosis in different patient cohorts (percentage and absolute numbers)^a

| Patient group | Sensitivity | Specificity | PPV | NPV |
|-----------------------------|--------------|---------------|--------------|---------------|
| Overall ^b | 73% (83/113) | 90% (498/552) | 61% (83/137) | 94% (498/528) |
| Solid organ transplantation | 94% (15/16) | 92% (89/97) | 65% (15/23) | 99% (89/90) |
| Intensive care unit | 79% (26/33) | 85% (176/206) | 46% (26/56) | 96% (176/183) |
| Respiratory diseases | 78% (25/32) | 91% (196/215) | 57% (25/44) | 97% (196/203) |
| Hematological malignancies | 67% (36/54) | 91% (126/139) | 73% (36/49) | 88% (126/144) |

Hoeningl J et al. Infect. 2012

Hoeningl M et al. Journal of Clinical Microbiology 2014

Pan Z et al J Med Microbiol. 2015

Heldt Curr Fungal Infect Rep 2017

Lateral Flow assay



Alastruey-Izquierdo TIMM 2017



Medical Mycology, 2014, 52, 647-652
doi: 10.1093/mmy/nyu019
Advance Access Publication Date: 17 June 2014
Short Communication



Short Communication

Serum and urine galactomannan testing for screening in patients with hematological malignancies

Wiebke Duettmann^{1,2}, Christoph Koidl³, Katharina Troppan², Katharina Seeber¹, Walter Buzina³, Albert Wöller², Jasmin Wagner¹, Robert Krause¹ and Martin Hoenigl^{1,4,*}

✓ Galactomannan in urine



ine

Urine Galactomannan-to-Creatinine Ratio for Detection of Invasive Aspergillosis in Patients with Hematological Malignancies

0.1)

Frederike M. J. Reischies,^a Reinhard B. Raggam,^b Juergen Prattes,^a Robert Krause,^a Susanne Eigl,^c Agnes List,^d Franz Quehenberger,^e Volker Strenger,^f Albert Wöller,^d Martin Hoenigl^{a,c,g}

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Duettmann et al. Med Mycol. 2014;52.
Reischies et al. J Clin Microbiol. 2016c;54(3):771-4.
Dufresne SF, et al. PLoS One. 2012;7(8)
Heldt Curr Fungal Infect Rep 2017

OPEN ACCESS Freely available online



Detection of Urinary Excreted Fungal Galactomannan-like Antigens for Diagnosis of Invasive Aspergillosis

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¹ Johns Hopkins University School of Medicine, Baltimore, Maryland, United States of America, ² The Sidney Kimmel Comprehensive Cancer Center, Johns Hopkins Medical Institution, Baltimore, Maryland, United States of America, ³ Albert Einstein College of Medicine, Bronx, New York, United States of America, ⁴ University of Texas Health Science Center and South Texas Veterans Healthcare System, San Antonio, Texas, United States of America, ⁵ Department de Microbiologia en Immunologia, Hospital de l'Esperança, Montornès (Barcelona), Canada, ⁶ McGill Medical University, Stomping People's Republic of China

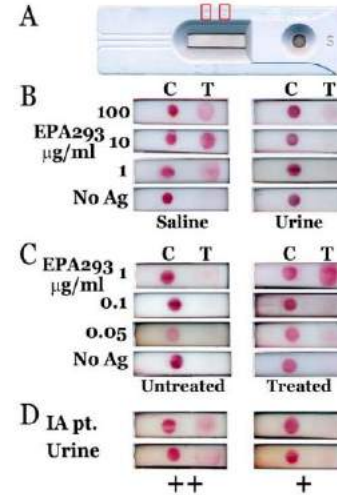
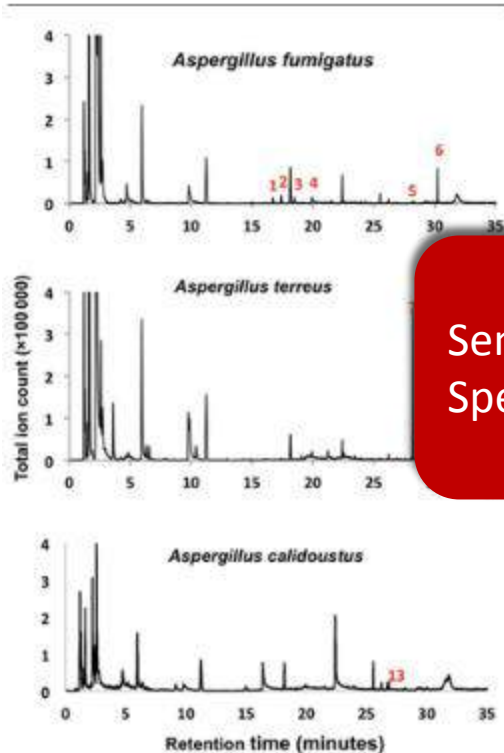


Figure 5. MAb476-based Lateral flow immunochromatographic assay device (LFD) detects Gal/antigen in simulated and human urine samples. A, appearance of the assembled LFD; subsequent panels show scanned images of the reaction window. B, Comparison of urine and normal saline (NS) as diluent for EPA293 in LFD. C, Detection of EPA293 in simulated samples after concentration (5-10 folds, 3 kDa MWCO) and desalting (7 kDa MWCO), compared to no treatment. D, Detection of Gal/antigen in processed clinical urine samples; 2 patients have intermediate (++) positive signal and 1 have weak (+) positive signal.
doi:10.1371/journal.pone.0042736.g005

Volatile organic compounds



Sensitivity: 94%
Specificity: 93%

Table 3. Breath Aspergillus Metabolite Signature by the Reference Standard and Test Parameters

| Parameter | Invasive Aspergillosis ^a | Other Pneumonia | Total Patients |
|--|-------------------------------------|-----------------|----------------|
| <i>Aspergillus</i> metabolite signature ^b + | 32 | 2 | 34 |
| <i>Aspergillus</i> metabolite signature – | 2 ^c | 28 | 30 |
| Total patients | 34 | 30 | 64 |
| Test parameters | | | |
| Sensitivity (95% CI) | 0.94 (.81–.98) | | |
| Specificity (95% CI) | 0.93 (.79–.98) | | |
| Positive likelihood ratio (95% CI) | 14.1 (3.69–54.0) | | |
| Negative likelihood ratio (95% CI) | 0.063 (.02–.24) | | |

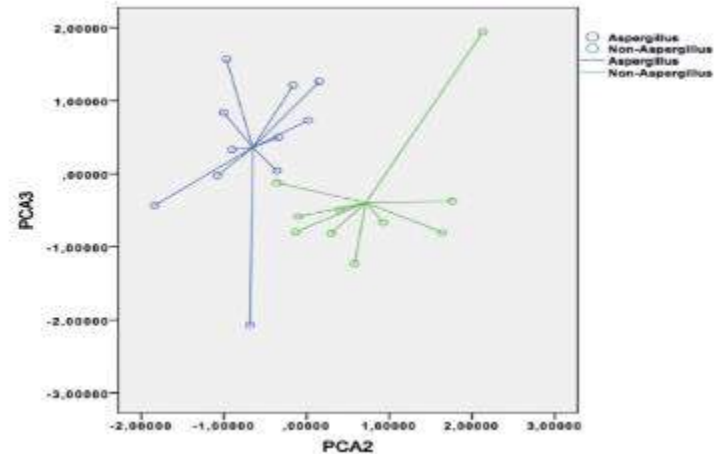
Electronic nose technology in COPD

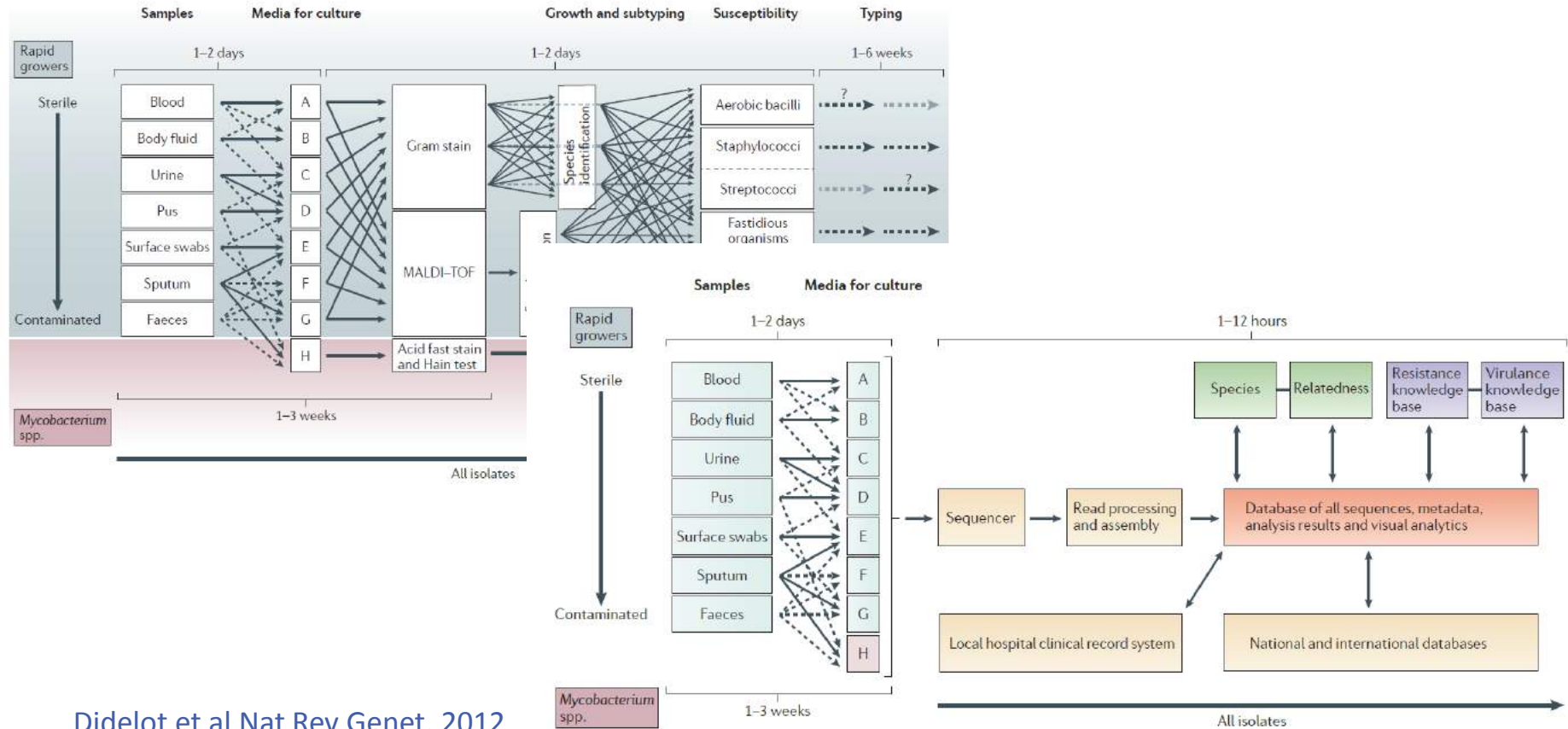
11 COPD
A.fumigatus +



10 COPD
A.fumigatus
-

Sensor data reduced to 2 PC
Cross-validity accuracy 90.5%
Sensitivity: 91%
Specificity: 90%





Application of next generation sequencing in clinical microbiology and infection prevention

Ruud H. Deurenberg², Erik Bathoorn^{2,1}, Monika A. Chlebowicz^{2,1}, Natacha Couto^{2,1}, Mithila Ferdous^{2,1}, Silvia Garcia-Cobos^{2,1}, Anna M.D. Kooistra-Smid^{2,b,1}, Erwin C. Raangs^{2,1}, Sigrid Rosema^{2,1}, Alida C.M. Veloo^{2,1}, Kai Zhou^{c,1}, Alexander W. Friedrich³, John W.A. Rossen^{2,4}

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R.H. Deurenberg et al. / Journal of Biotechnology 243 (2017) 16–24

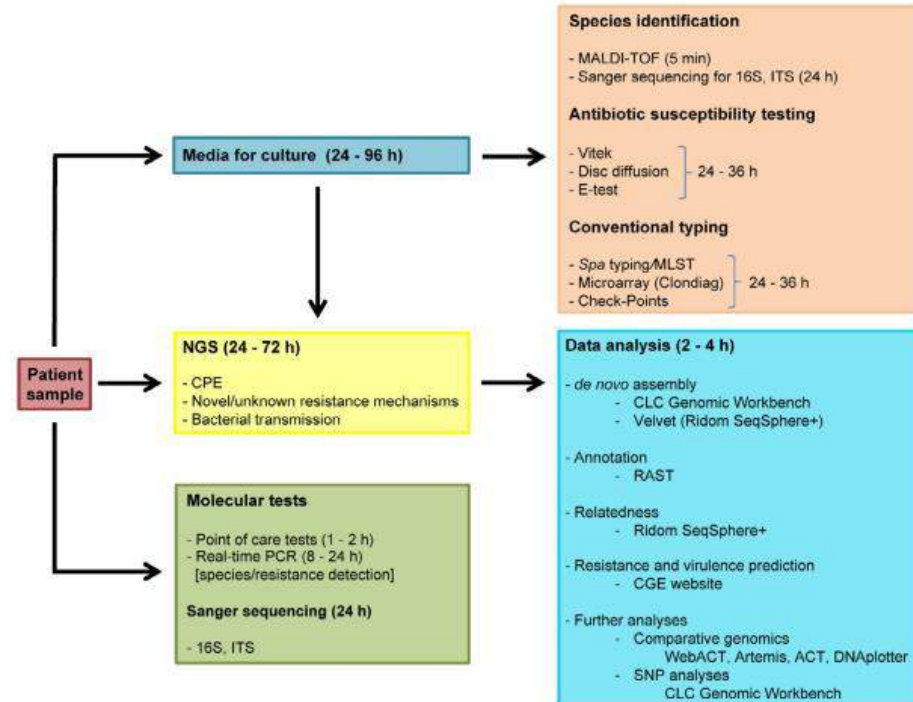
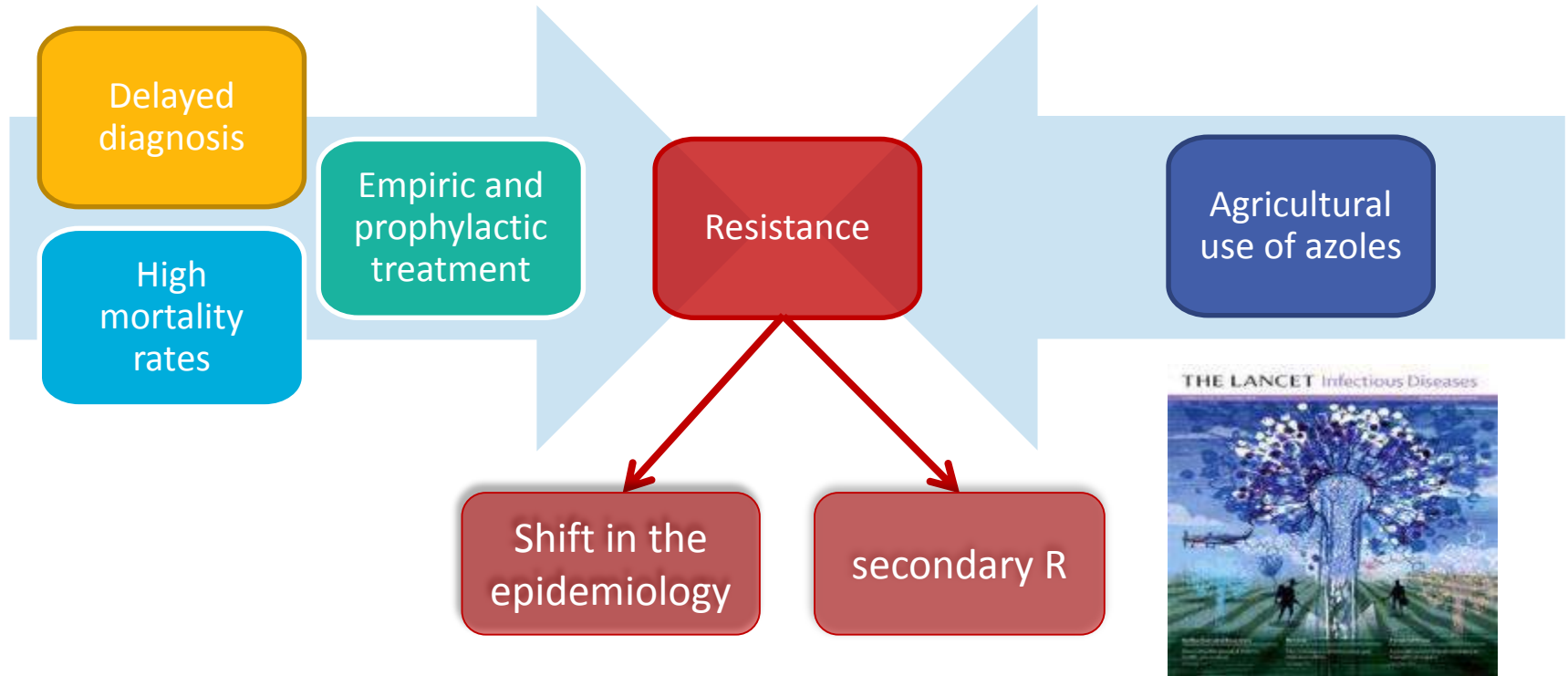


Fig. 1. A schematic overview of the general workflow of diagnostic procedures including NGS in our laboratory.

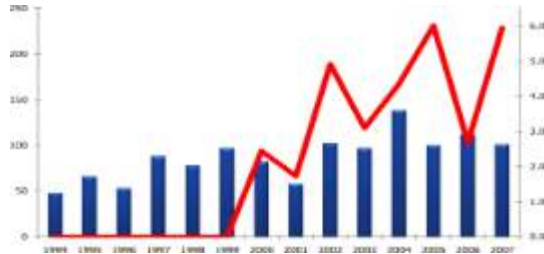
AF resistance



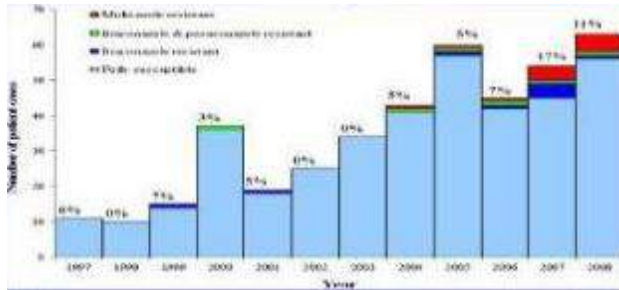
AF resistance *Aspergillus*



Alastruey-Izquierdo TIMM 2017



Snelders et al., PLOS 2008 Vol. 5:11



Howard et al., EID 2009, Vol. 15, No. 7

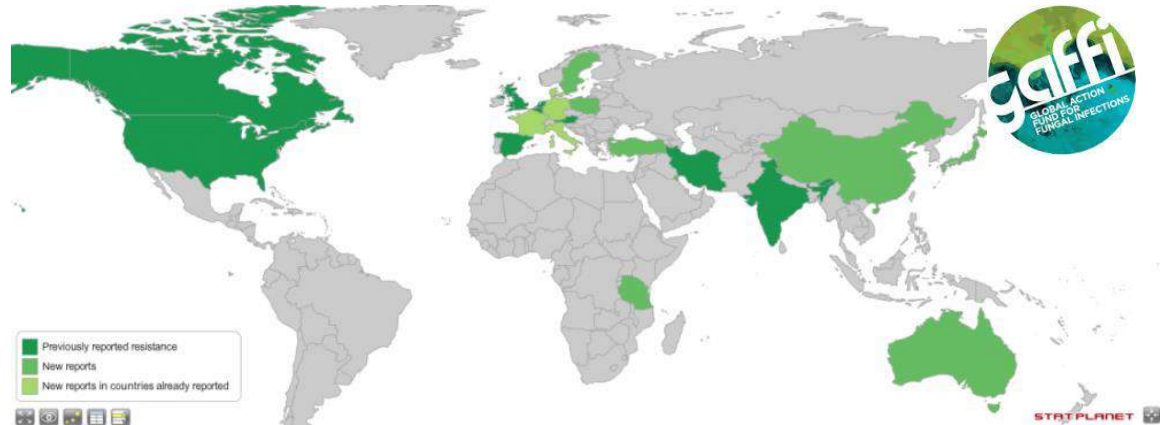
Azole antifungal drug resistance in *Aspergillus fumigatus* & resistance mechanisms REVIEW



Figure 2. *Aspergillus fumigatus* cyp51A gene depicting the promoter region and mutation hot spots conferring resistance to azole antifungals.

Chowdhari Future Micro 2015

Secondary resistance



<http://www.gaffi.org/uk-calls-for-agricultural-fungicide-restraint-to-reduce-azole-resistance-in-aspergillus/>

Aspergillus cryptic species



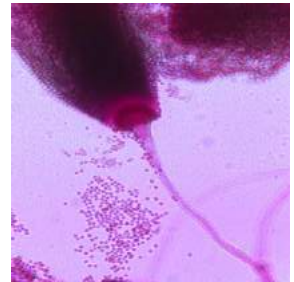
Alastruey-Izquierdo TIMM 2017

Table 1. *Aspergillus* species in epidemiological surveys from Spain and the U. S.²²

| Species | Section | Transnet | | FILPOP | |
|--------------------------|--------------|------------|------|------------|------|
| | | N isolates | % | N isolates | % |
| <i>A. fumigatus</i> | Fumigati | 139 | 63.8 | 156 | 56.1 |
| <i>A. lentulus</i> | Fumigati | 4 | 1.8 | 3 | 1.1 |
| <i>A. udagawae</i> | Fumigati | 3 | 1.4 | 0 | 0.0 |
| <i>N. pseudofischeri</i> | Fumigati | 1 | 0.5 | 1 | 0.4 |
| <i>A. viridinutans</i> | Fumigati | 0 | 0.0 | 1 | 0.4 |
| <i>A. fumigatiafinis</i> | Fumigati | 0 | 0.0 | 1 | 0.4 |
| <i>A. flavus</i> | Flavi | 29 | 13.3 | 27 | 9.7 |
| <i>A. alliaceus</i> | Flavi | 0 | 0.0 | 3 | 1.1 |
| <i>A. terreus</i> | Terrei | 11 | 5.0 | 26 | 9.4 |
| <i>A. carneus</i> | Terrei | 0 | 0.0 | 1 | 0.4 |
| <i>A. tubingensis</i> | Nigri | 6 | 2.8 | 22 | 7.9 |
| <i>A. niger</i> | Nigri | 13 | 6.0 | 21 | 7.6 |
| <i>A. calidoustus</i> | Usti | 6 | 2.8 | 4 | 1.4 |
| <i>A. insuetus</i> | Usti | 0 | 0.0 | 1 | 0.4 |
| <i>A. keveii</i> | Usti | 0 | 0.0 | 1 | 0.4 |
| <i>A. sydowii</i> | Versicolores | 2 | 0.9 | 1 | 0.4 |
| <i>A. versicolor</i> | Versicolores | 3 | 1.4 | 0 | 0.0 |
| <i>E. quadrilineata</i> | Nidulantes | 1 | 0.5 | 0 | 0.0 |
| <i>A. nidulans</i> | Nidulantes | 0 | 0.0 | 8 | 2.9 |
| <i>A. westerdijkiae</i> | Circumdati | 0 | 0.0 | 1 | 0.4 |
| Total | | 218 | 100 | 278 | 100 |

11%

15%



Balajee et al, JCM 2009

Alastruey-Izquierdo et al. AAC 2013

Alastruey-Izquierdo et al. ANYAS 2012

Aspergillus cryptic species



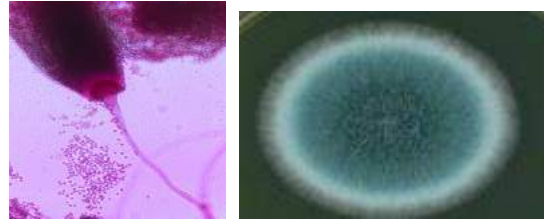
Alastruey-Izquierdo TIMM 2017

| | n | AMB | ICZ | VCZ | PCZ | CPF | MCF | ANF |
|---------------------------|----|--------------|-------------|-------------|-------------|-------|------|------|
| <i>A. lentulus</i> | 26 | 3 | 2.3 | 3.4 | 0.23 | 1.6 | 0.1 | 0.1 |
| <i>N. hiratsukae</i> | 9 | 1.7 | 0.9 | 1.1 | 0.16 | 0.11 | 0.03 | 0.03 |
| <i>N. pseudofischerii</i> | 6 | 0.25 | 4 | 2.51 | 0.22 | 0.86 | 0.03 | 0.03 |
| <i>A. fumigatiaffinis</i> | 6 | 4.8 | 5 | 3.1 | 0.4 | 0.22 | 0,03 | 0,03 |
| <i>N. udagawae</i> | 5 | 2 | 0.6 | 2.3 | 0.25 | 0.3 | 0.03 | 0.03 |
| <i>A. viridinutans</i> | 3 | 0,7 | 16 | 4 | 0,25 | 5,66 | 0,06 | 0,09 |
| <i>A. tubingensis</i> | 22 | 0.11 | 0.42 | 0.76 | 0.09 | 0.3 | 0.05 | 0.03 |
| <i>A. calidoustus</i> | 19 | 0.9 | 8.6 | 6.2 | 6.8 | 0.5 | 0.04 | 0.04 |
| <i>A. insuetus</i> | 2 | 0.7 | 11.3 | 8 | 2.8 | 5.6 | 1.4 | 0.9 |
| <i>A. keveii</i> | 1 | 0,25 | 16 | 16 | 16 | 16 | 16 | 16 |
| <i>A. alliaceus</i> | 30 | 19.25 | 0.2 | 0.5 | 0.11 | 12.15 | 3.8 | 1.9 |

Screening for azole resistance in the lab



Alastruey-Izquierdo TIMM 2017



40 WT and 39 Azole R mutants
3 labs/2 readers
Good agreement

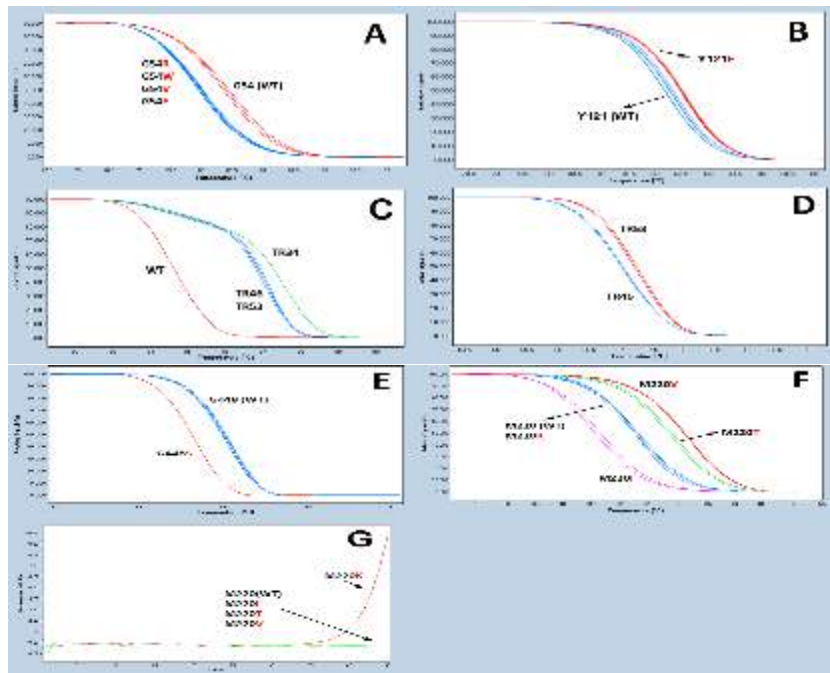


Sensitivity & specificity based on presence of *CYP51A* mutations & growth in >1 azole containing agar well

| Centre | Balis Plates - Observer 1 | | PV Plates - Observer 1 | | Balis Plates - Observer 2 | | PV Plates - Observer 2 | |
|--------------------|---------------------------|-------|------------------------|-------|---------------------------|-------|------------------------|-------|
| | Sens. | Spec. | Sens. | Spec. | Sens. | Spec. | Sens. | Spec. |
| ITC/VRC/PSC | | | | | | | | |
| Overall | 99% | 99% | 99% | 98% | 99% | 100% | 99% | 98% |
| ITC/VRC | | | | | | | | |
| Overall | 99% | 99% | 99% | 98% | 99% | 100% | 98% | 98% |
| ITC/PSC | | | | | | | | |
| Overall | 88% | 99% | 88% | 98% | 87% | 100% | 89% | 98% |
| VRC/PSC | | | | | | | | |
| Overall | 85% | 99% | 85% | 100% | 89% | 100% | 88% | 100% |

Meletiadis et al. ECCMID 2017 # P1750

Azole resistance detection



Bernal-Martinez AAC 2017



AsperGenius

Species multiplex

- *Aspergillus fumigatus*
- *Aspergillus terreus*
- *Aspergillus species*
- Internal Amplification Control (IAC)

Resistance multiplex

- L98H
- Tandem repeat 34
- T289A
- Y121F

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Azole resistance in *Aspergillus fumigatus* from bronchoalveolar lavage fluid samples of patients with chronic diseases

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Table 4. Mutations in the CYP51A gene of *A. fumigatus* detected directly from BAL fluid samples

| No. of samples | Amino acid substitutions caused by non-synonymous mutations | Polymorphisms caused by synonymous mutations | Culture/azole susceptibility |
|--|---|--|--|
| Samples carrying single or multiple non-synonymous mutations | | | |
| 1 | N425S | | <i>A. fumigatus</i> /susceptible |
| 1 | M172V ^a , F46Y ^a | | <i>A. fumigatus</i> /susceptible |
| 1 | L206P, L210P | G89G | negative/NA |
| 1 | K67E | G89G | <i>Penicillium</i> species/POS=1, VRC=4, ITC > 4 |
| 1 | Y107C | D70D, G89G | negative/NA |
| 1 | M220V^b | G89G | <i>A. fumigatus</i> /susceptible |
| 1 | N33D, K80E | G89G | ND/NA |
| 1 | F41S, E66G | D70D, G89G | ND/NA |
| 1 | P216L^c | G89G | negative/NA |
| Samples carrying synonymous mutations only | | | |
| 1 | | G89G, F495F | ND/NA |
| 4 | | D70D, G89G | 3 ND; 1 negative/NA |
| 1 | | V44V, G89G | negative/NA |
| 1 | | G89G, F165F | <i>A. fumigatus</i> /susceptible |
| 38 | | G89G | one resistant <i>A. fumigatus</i> isolate; POS=1, VRC=2, ITC > 4 |

Summary



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- ✓ No perfect diagnostic test nor diagnosis algorithm for IA
- ✓ Combination of tests
- ✓ Future promises: LFD / VOCs
- ✓ Azole resistance emerging
- ✓ Detection methods available
- ✓ Treatment options should be adapted to local situations
- ✓ AFST, epidemiological studies
- ✓ New antifungals

Thanks for your attention



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