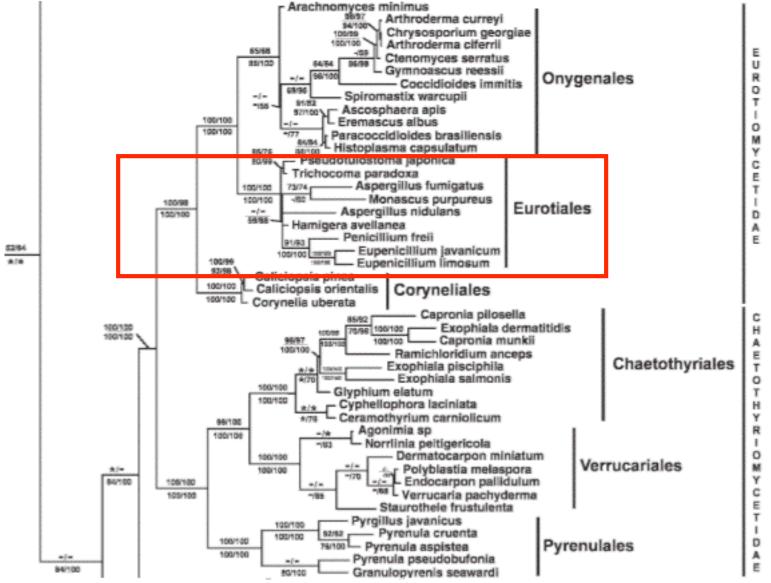
Regulation of cellular morphogenesis during development in *Aspergillus nidulans*

Steven D. Harris

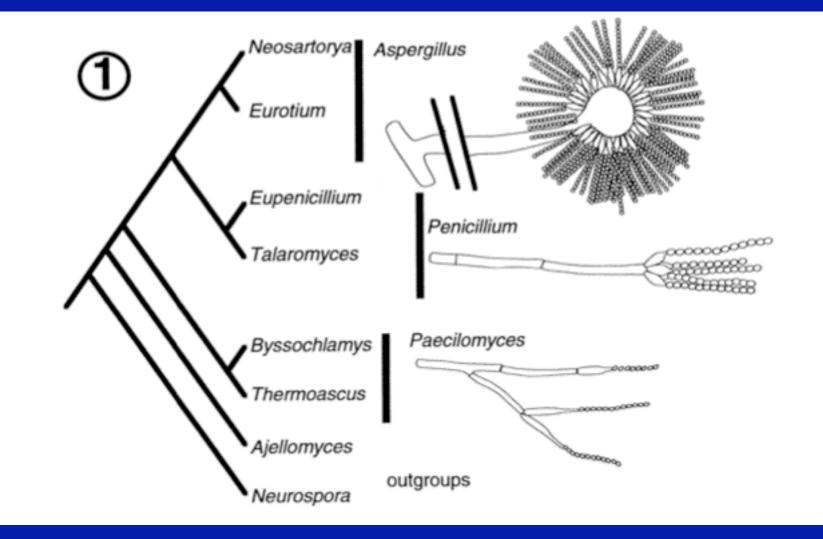
Center for Plant Science Innovation/Dept. Plant Pathology University of Nebraska Lincoln, NE

Evolution of conidiophore morphology in the Eurotiales

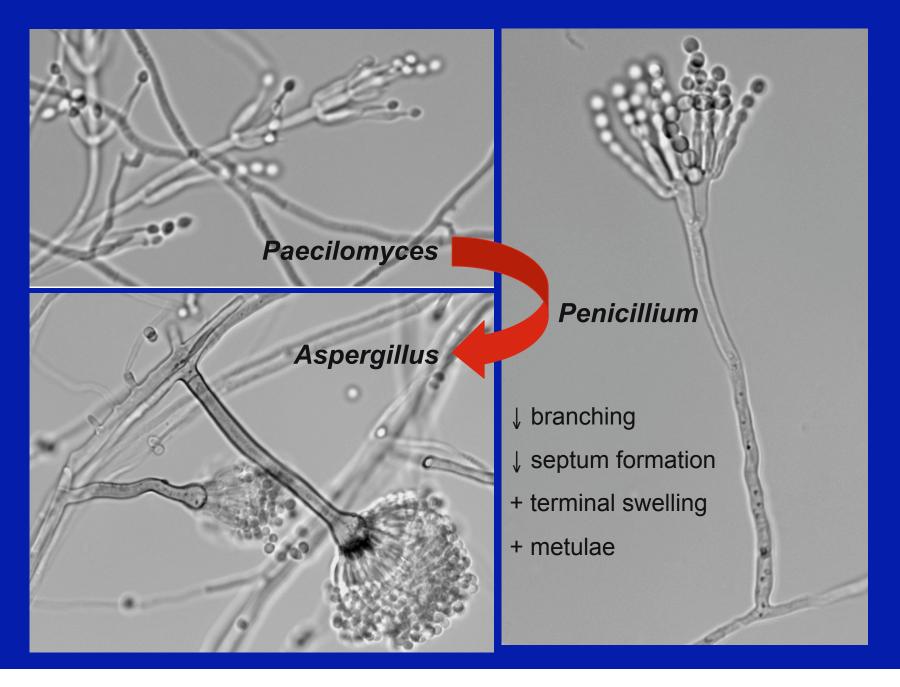


E υ R О M γ С Ε Т Ε S

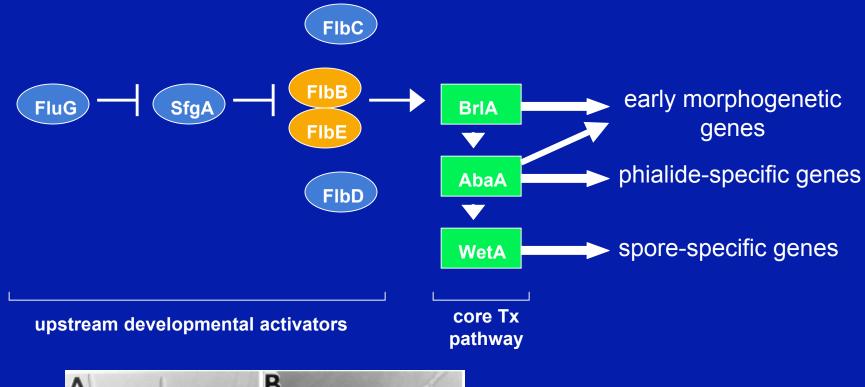
Evolution of conidiophore morphology in the Eurotiales

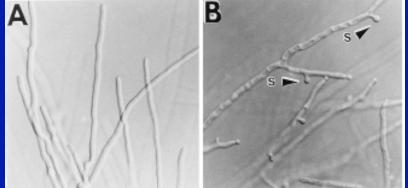


Increasing complexity of conidiophore architecture



Regulation of conidiophore development in A. nidulans





expression of *alcA*(p)::*brlA* drives formation of spores in submerged hyphae

How are cell cycle and morphogenetic functions integrated within this regulatory network?

Potential factors underlying the evolution of conidiophore morphology

- 1. Cell cycle regulation Wee1 tyrosine kinase; Ace2/Swi5 C2H2 transcription factor
- 2. Spatial regulation of cellular morphogenesis Cdc42 GTPase module
- Integration with UDAs and core transcriptional regulatory network
 BrIA; AbaA

Why?

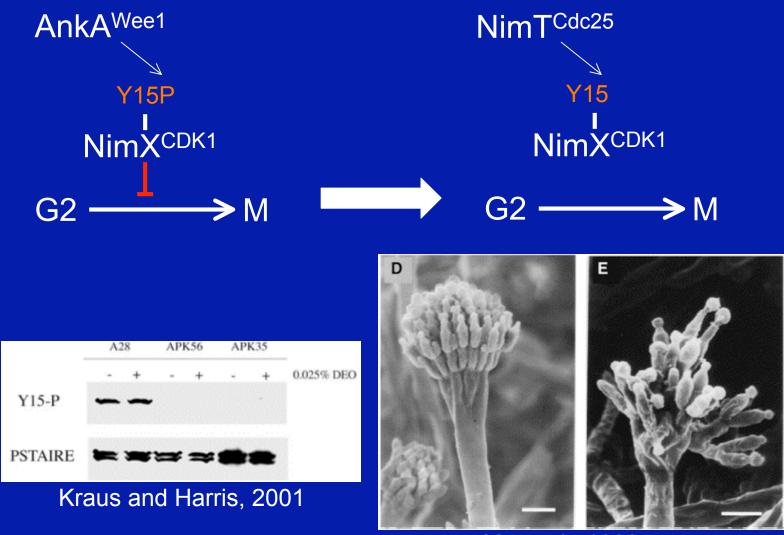
-- improved understanding of conidiophore morphogenesis

-- understand the molecular events involved in the evolution of conidiophore morphology

-- identification of potential strategies for the control of sporulation

Progress →

Cell cycle regulation: Wee1

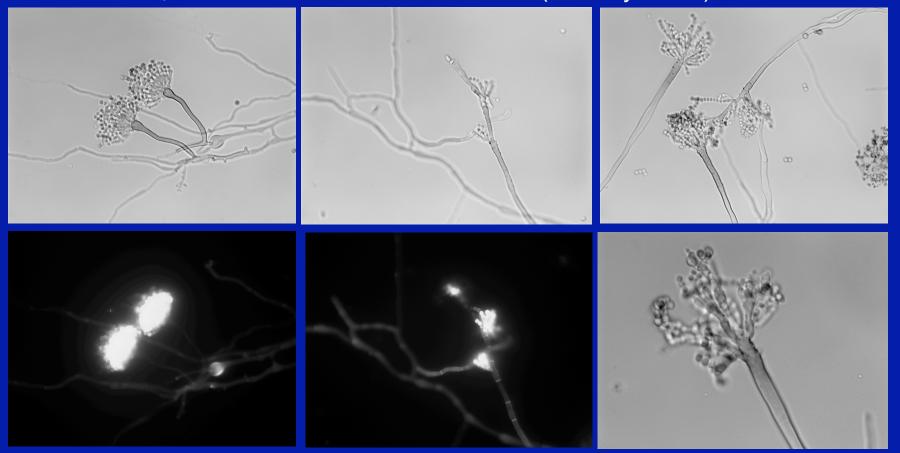


Ye et al., 1999

Cell cycle regulation: Wee1

wildtype

ankA2 (formerly sntA1)



Loss of Wee1 function (i.e., NimX^{CDK1}Y15 Pn) causes;

septum formation in stalk, loss of vesicle, spurious branch formation, defective hyphal → budding transition

Ace2 and Swi5 = paralogous C2H2 transcription factors in yeast

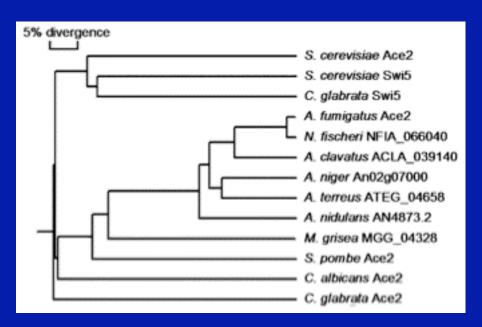
Ace2 → septum degradation following cytokinesis Swi5 → transcription of genes expressed at M/G1 transition and in G1

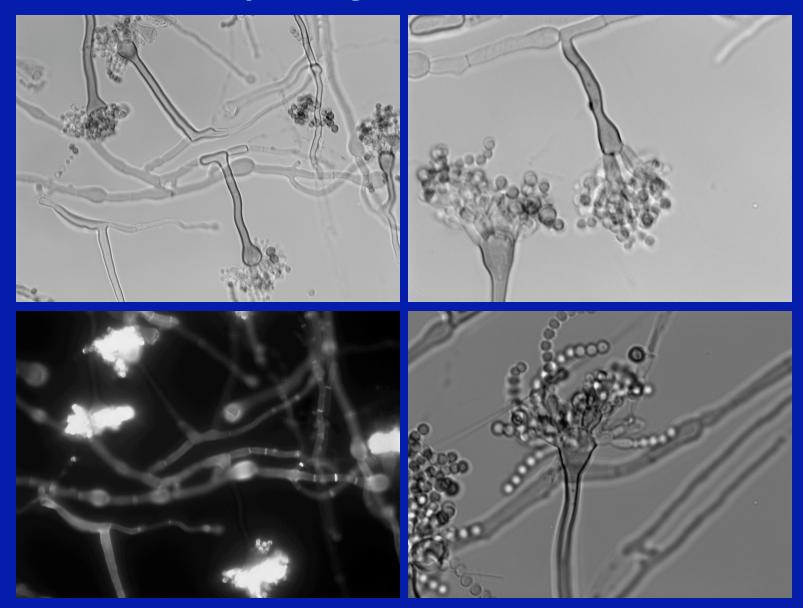
Afum Ace2 (Afu3g11250) required for phialide formation, cell wall architecture, and virulence (Ejzykowicz et al., 2009)

ANID_04873.1

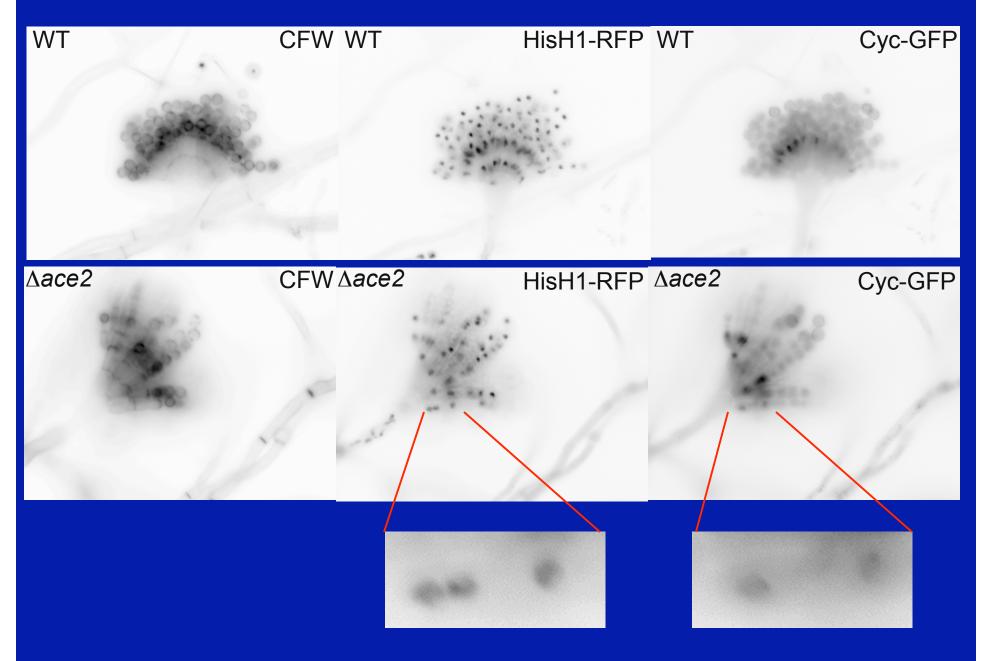
51%ID/71%S over 781 AA – Afum 55%ID/65%S over 110 AA – Swi5

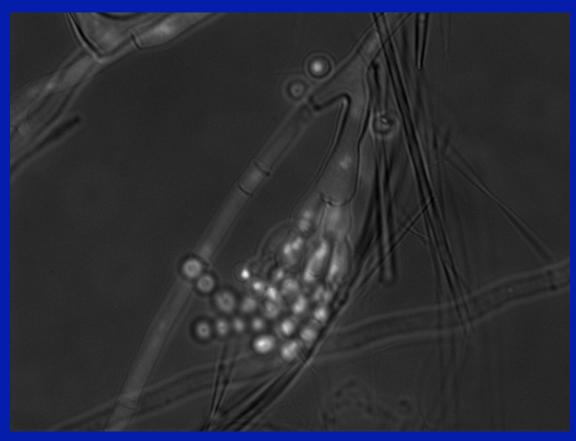
generated ∆*ace2::pyroA* mutants and screened for conidiation defects





normal stalk + vesicle; defective transition from vesicle to metulae (hyphal -> budding)



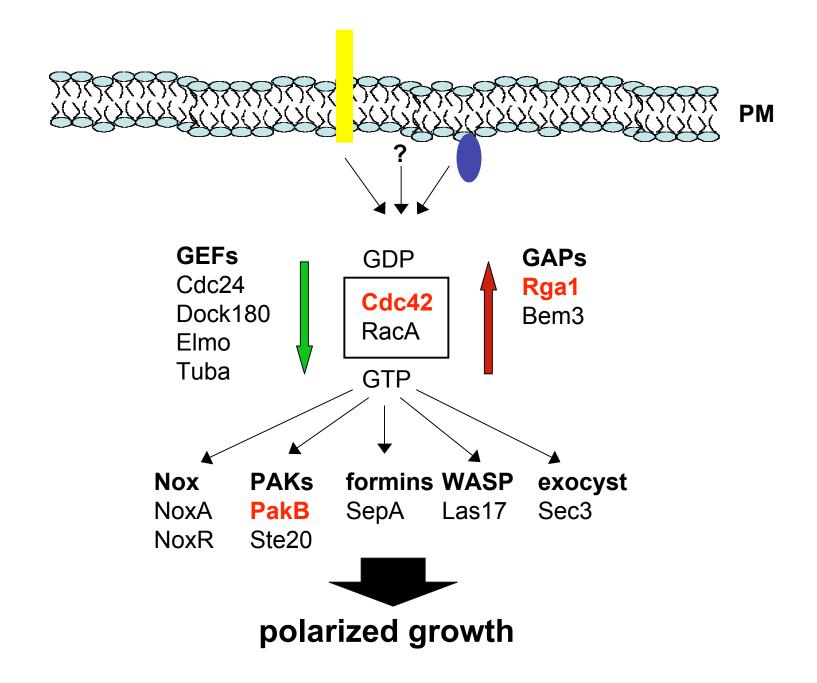


Ace2/Swi5 is required for orderly transition from vesicle → metulae

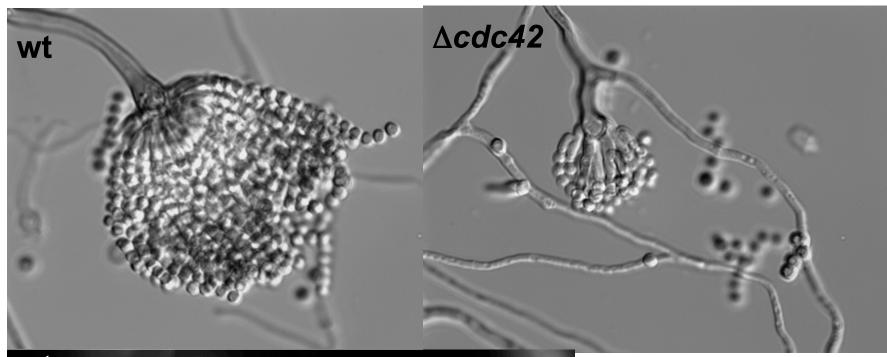
absence results in multinucleate hybrid cells that appear to undergo asynchronous mitosis

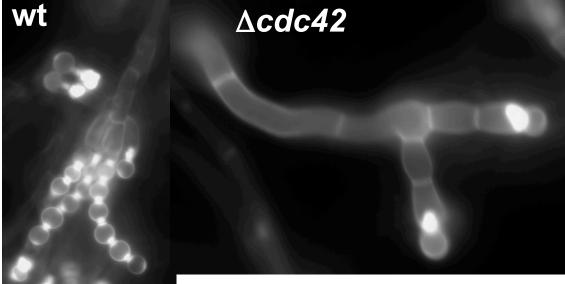
coordinated cell cycle regulation underlies morphological transitions

Cdc42/Rac GTPase module



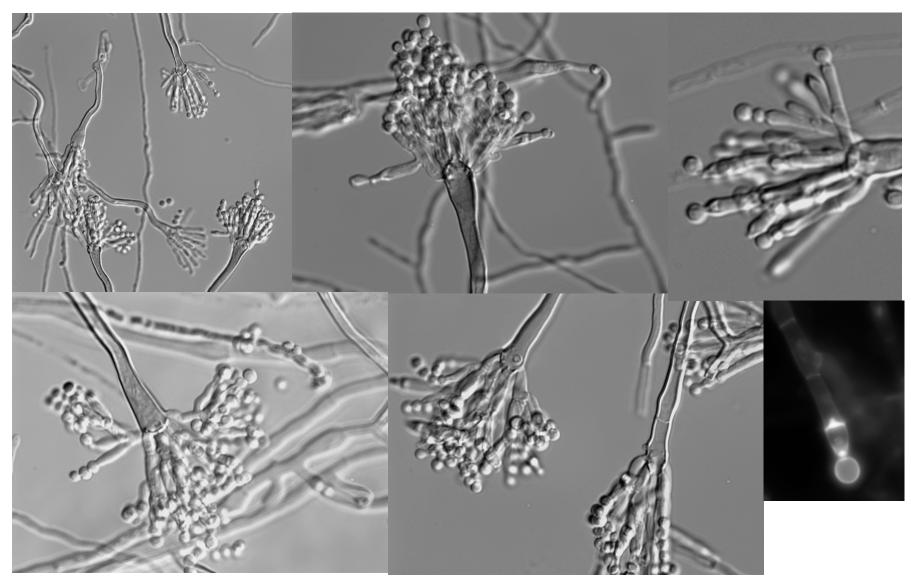
Conidiophore development: Cdc42





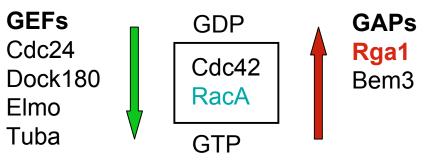
shorter conidiophores wider cells abnormal phialidespore junctions

Conidiophore development: Rga1



 $\Delta rga1$ -- no obvious vesicles; elongated metulae and phialides; random branching of metulae

Conidiophore development: Rga1

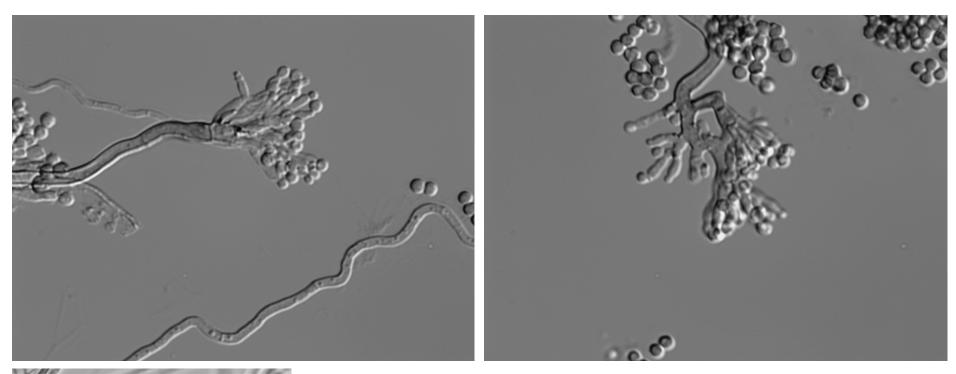


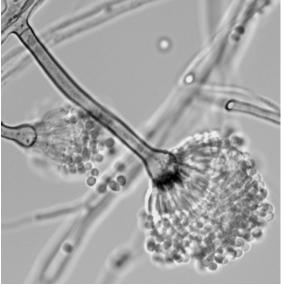
Does the $\Delta rga1$ phenotypes reflect activation of Cdc42 or Rac1?



Deletion of *cdc42* suppresses $\Delta rga1$ conidiophore defects. Thus, at least during conidiation, the primary target of Rga1 is Cdc42 (and $\Delta rga1$ defects are due to abnormally active Cdc42)

Conidiophore development: PakB





PakB;

suppresses conidiophore branching, formation of vesicle

Cdc42/Rac GTPase module

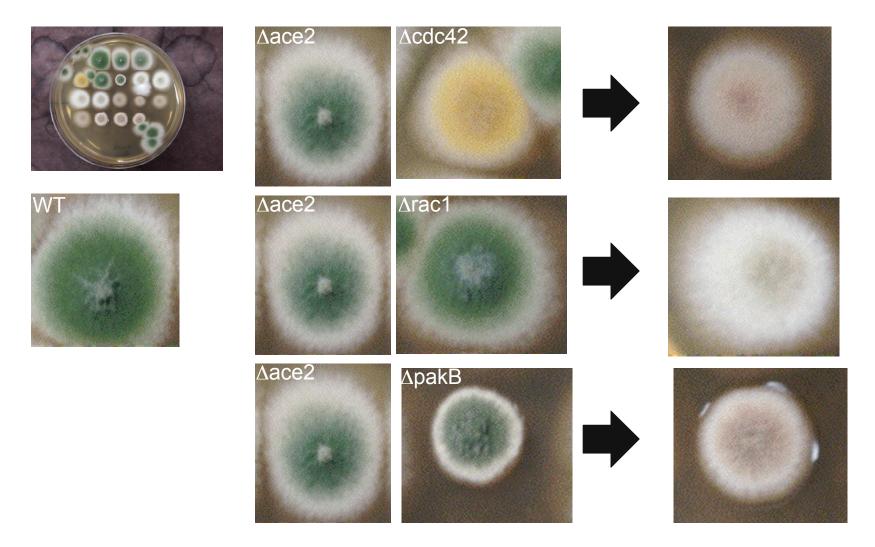
Rga1 and PakB suppress lateral branching during stalk formation --- proper spatial control of Cdc42 activation confines growth to tip and enables vesicle formation?

Rga1 is required for normal transition from hyphal to budding growth --- Cdc42 activity must be down-regulated to enable budding of metulae from vesicle?

Do cell cycle regulation and the Cdc42/Rac GTPase modules function in the same pathway?

Test double mutants for epistasis

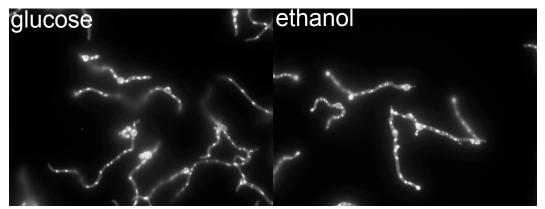
Genetic interaction analysis



Enhanced conidiation defects imply that Ace2/Swi5 and the Cdc42/Rac1 GTPase module act via different pathways to regulate development

Integration of cell cycle functions and Cdc42/Rac GTPase module with the core transcription pathway (BrIA)

Forced expression of *brlA* triggers formation of conidia (Adams and Timberlake, 1988)



alcA(p)::brlA

RNA-Seq was used to examine transcript profiles following *brlA* induction;

cell cycle

 nimX (CDK1)
 ↑2.5x

 ankA (Wee1)
 ↑2.2x

 nimE (G2 cyclin)
 ↑5.4x

 cln1 (G1 cyclin)
 ↑2.2x

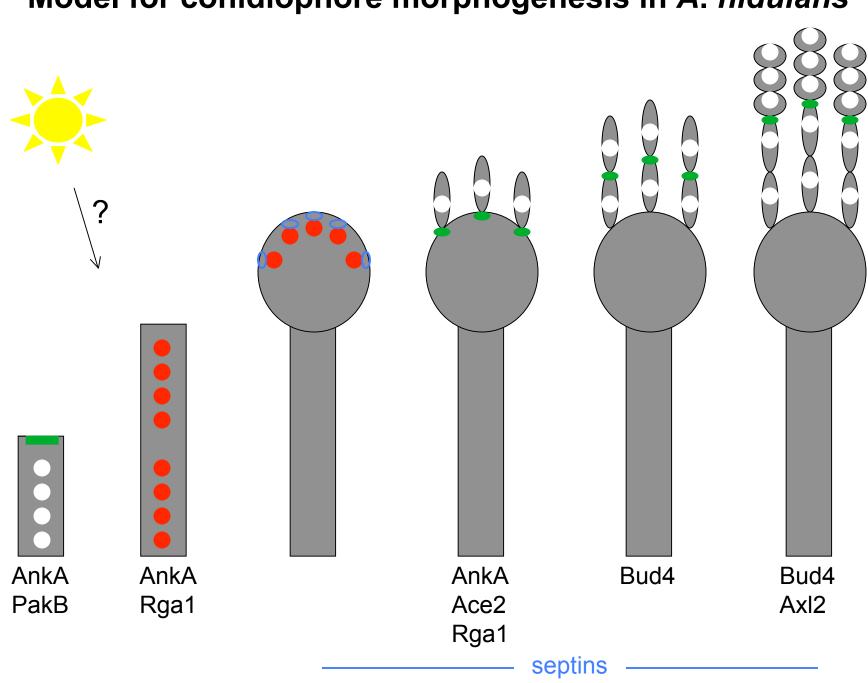
 pclA (G1 cyclin)
 ↑4.9x

 ace2/swi5
 ↑3.8x

Suggests that expression of cell cycle functions is regulated by BrIA→AbaA

stronger induction in response to *abaA* induction

Cdc42/Rac1 GTPase module – no response; post-transcriptional?



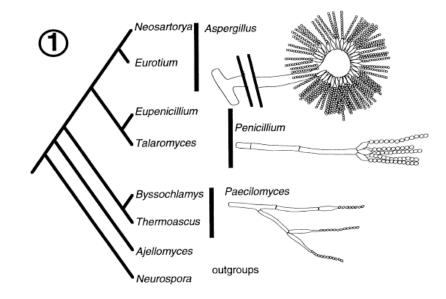
Model for conidiophore morphogenesis in A. nidulans

Evolution of conidiophore morphology

- -- suppression of branching and septum formation in stalk
- -- termination of polarized growth to form vesicle
- -- switch to budding growth

Targets of selection?

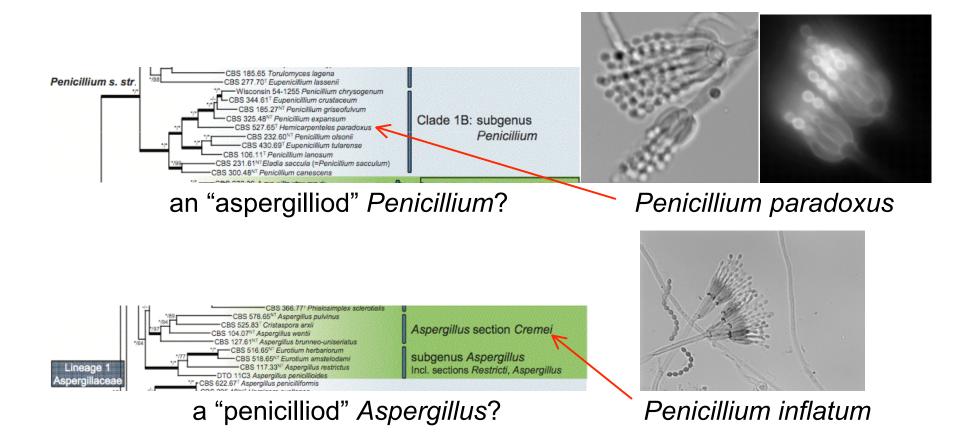
expression of *ankA*, *ace2/swi5*, cyclins, or other cell cycle regulators? post-transcriptional control of Cdc42 GTPase activity (phosphorylation of GEFs, GAPs)?



Evolution of conidiophore morphology

Test by;

1.Comparative gene expression analysis (e.g., *A. nidulans* vs. *P. chrysogenum*) during staged conidiation2.Examine morphologically aberrant/transitional species



Aleks Virag Haoyu Si Bill Rittenour Brad Downs Rennes Nassrat Lakshmi Yerra



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