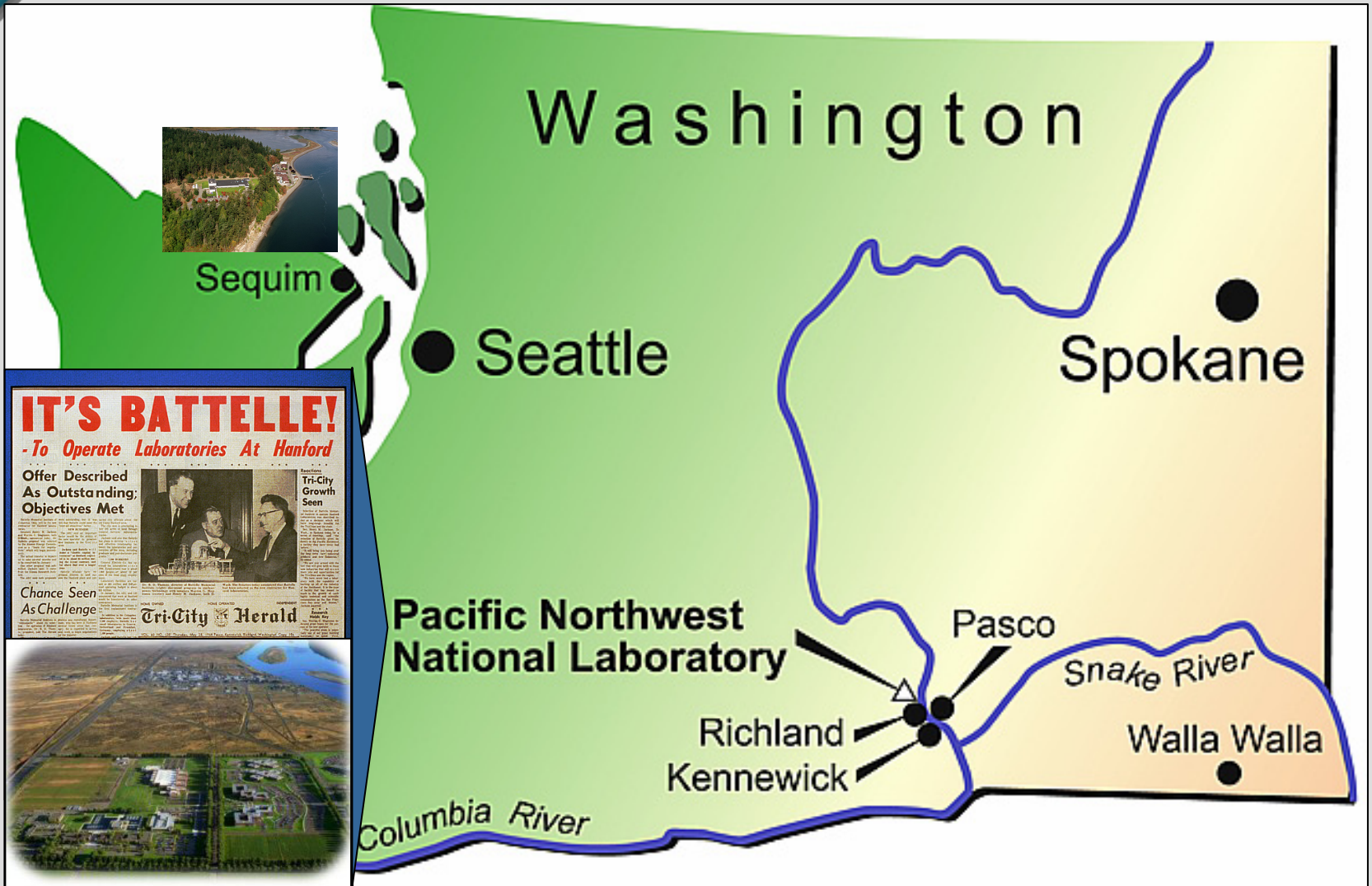


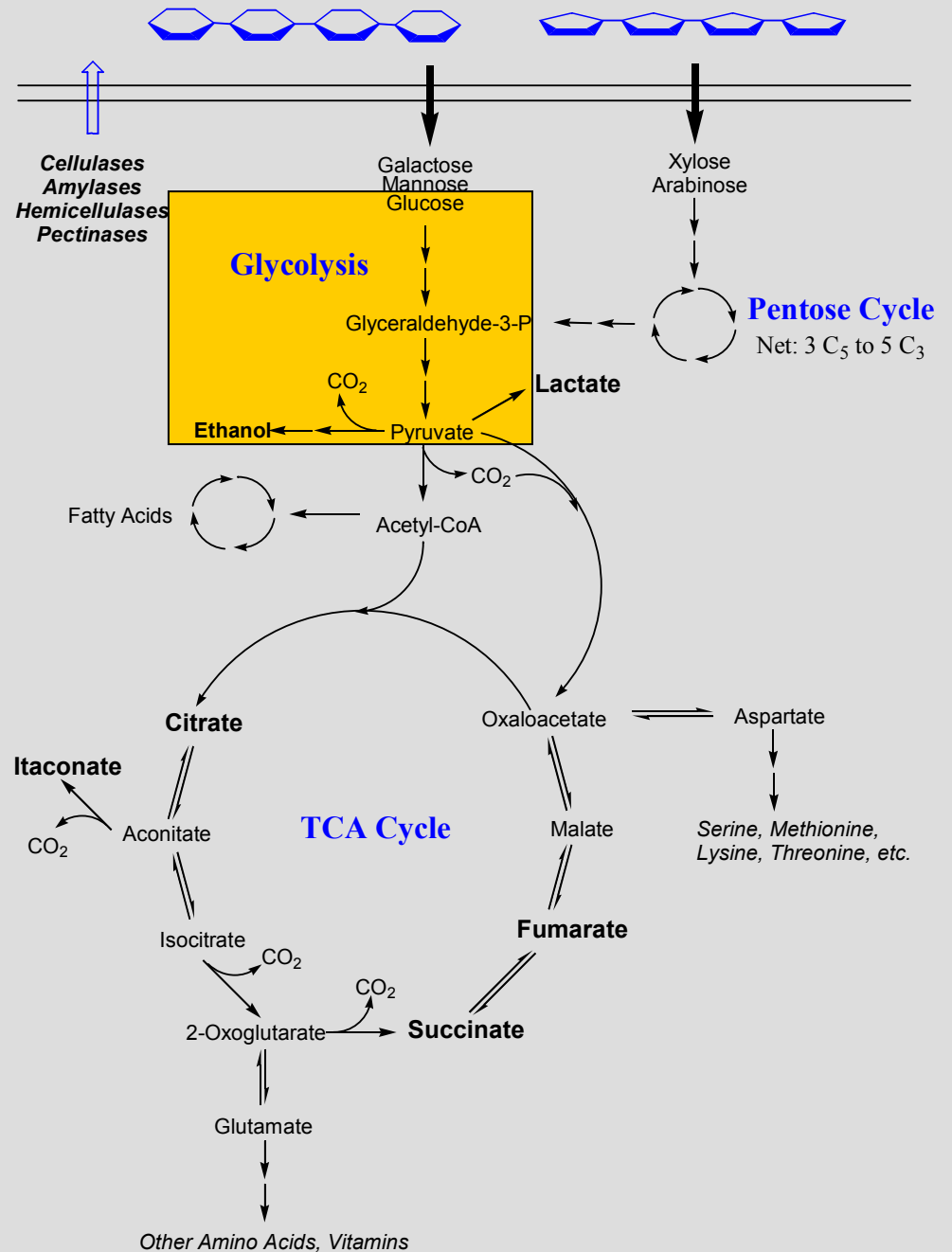
What's new in the *Aspergillus niger* genome?

Scott E. Baker
Pacific Northwest National Laboratory
Advances Against Aspergillosis 2006
Athens, Greece
February 25, 2006



Filamentous Fungi

- Nature's versatile biomass conversion factories
- 100's of enzymes for plant polymer hydrolysis
- Utilize both C₆ and C₅ sugars
- Produce 10s to 100s of grams/liter enzyme protein, organic acids



Current and **future** routes to fuels and chemicals

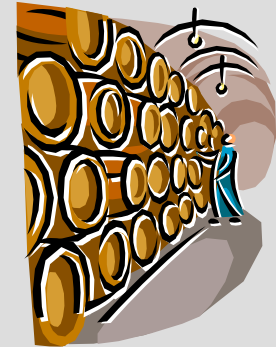
Petroleum products



Petroleum



Petroleum refinery



Products: Fuels and chemicals

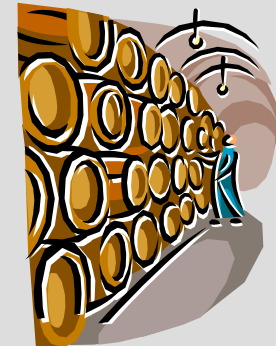
Biobased products



**Complex biomass:
Agricultural products
and “waste”**

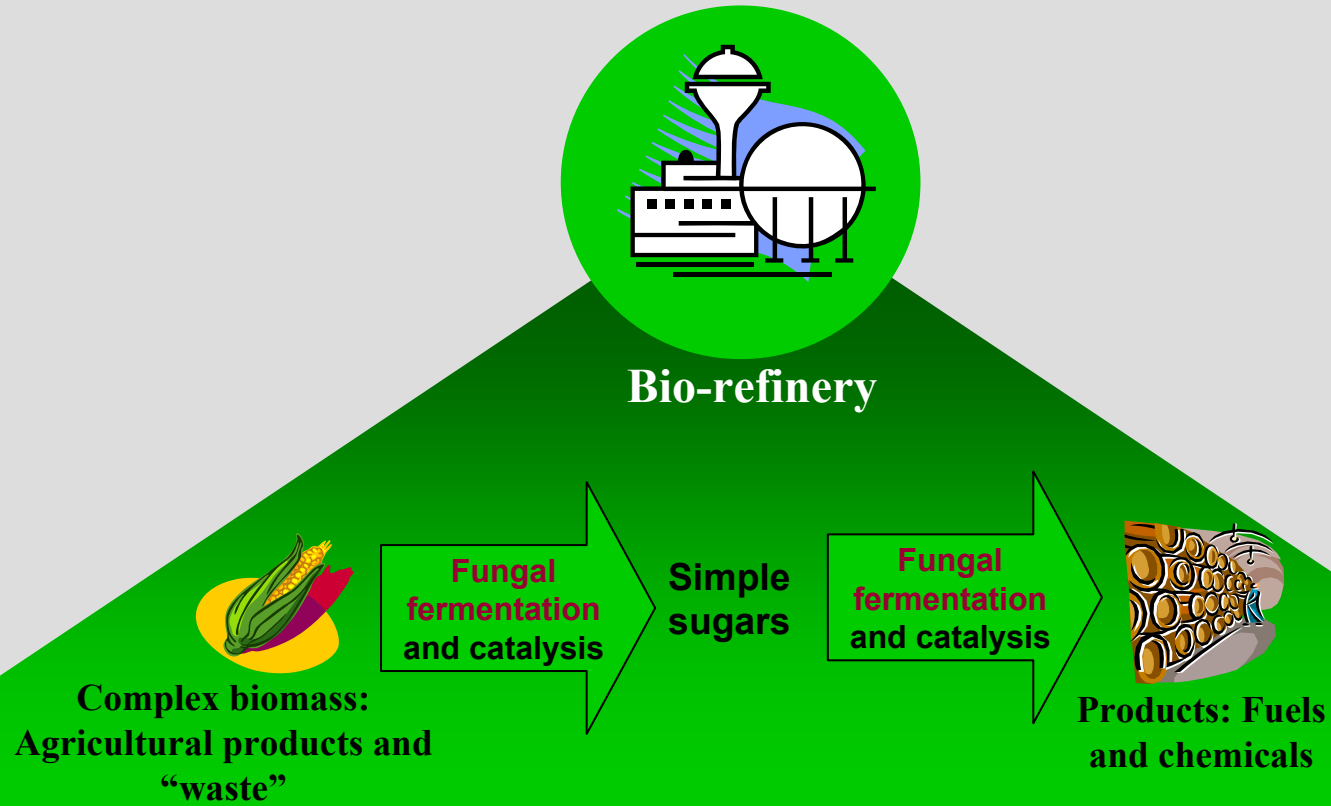


Bio-refinery



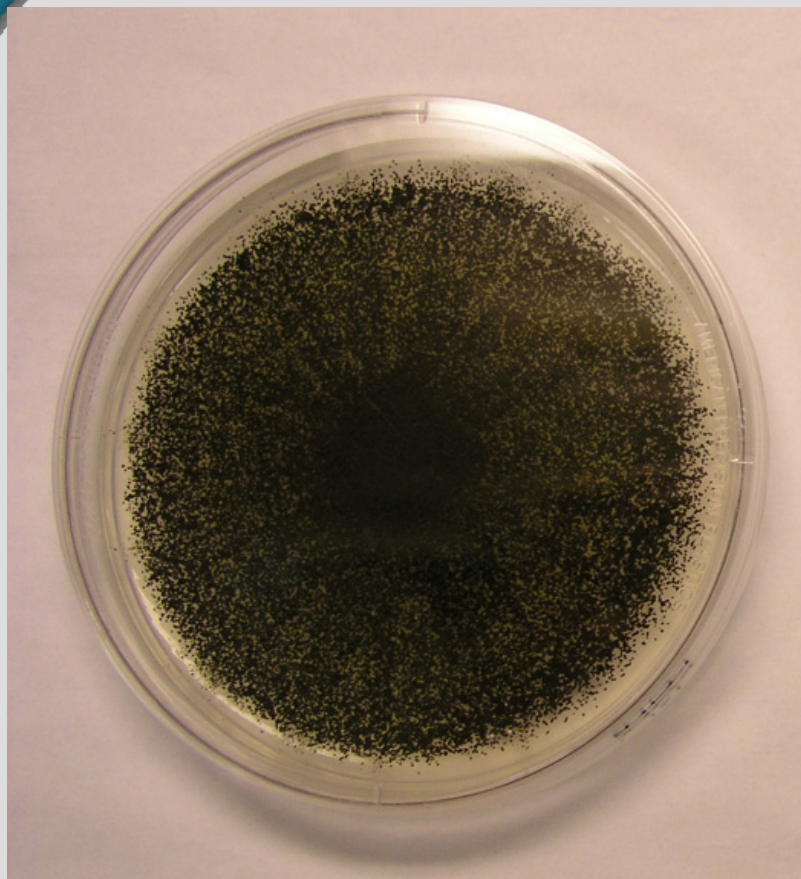
Products: Fuels and chemicals

Filamentous fungi inside the Bio-refinery

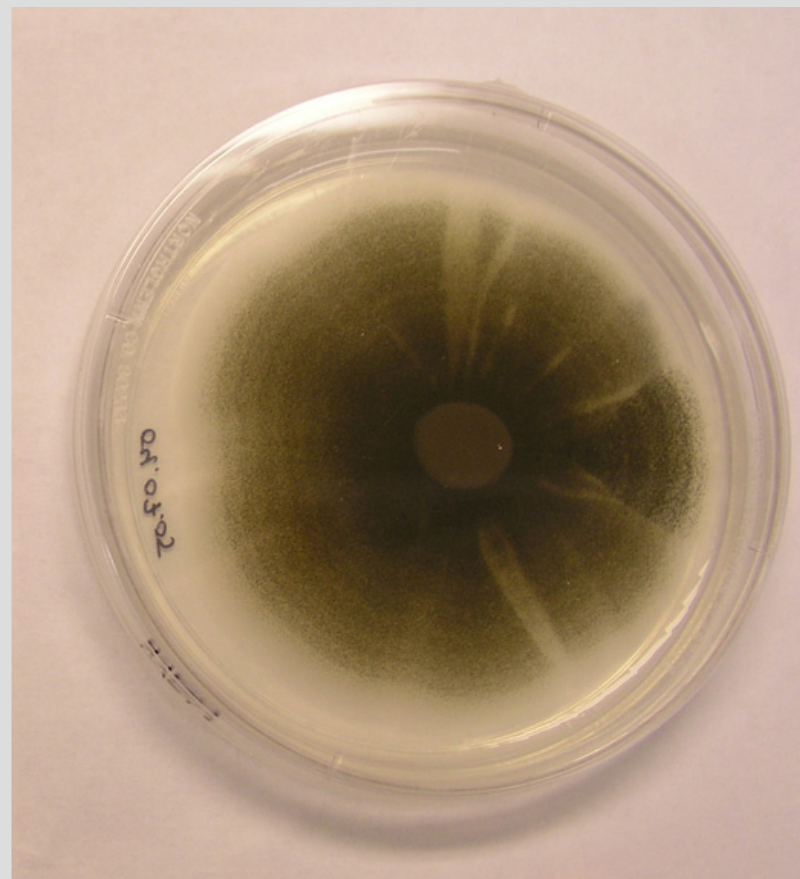


Why a public *Aspergillus niger* genome project?

- ▶ A bioprocess organism
 - First citric acid process reported in 1917 with wildtype ATCC 1015 *Aspergillus niger* strain
 - Highly efficient fermentation of glucose to citric acid
- ▶ A protein production organism
 - Source of important enzymes
 - Industrial protein producer
- ▶ Sequenced twice by industry
 - Public access to sequence with restriction
- ▶ Large economic footprint
- ▶ Pathogen of immunocompromised humans



Wild-type (ATCC 9029)

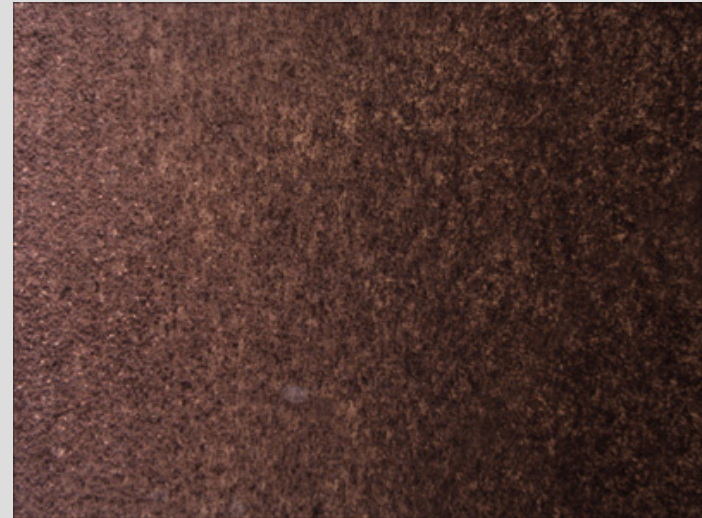
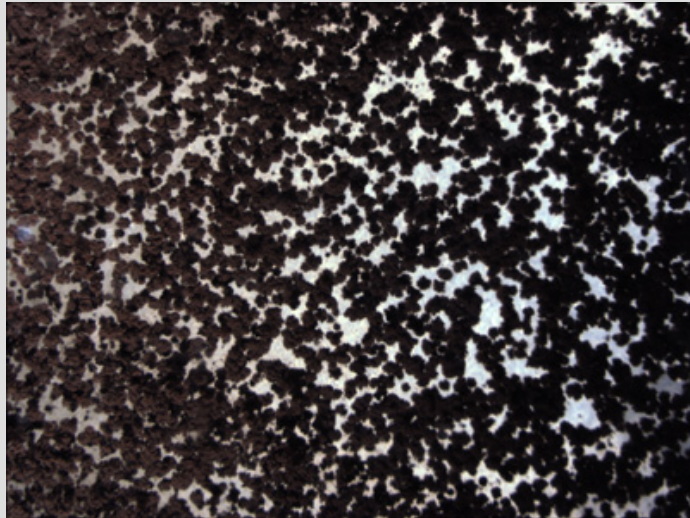


NRRL 3122

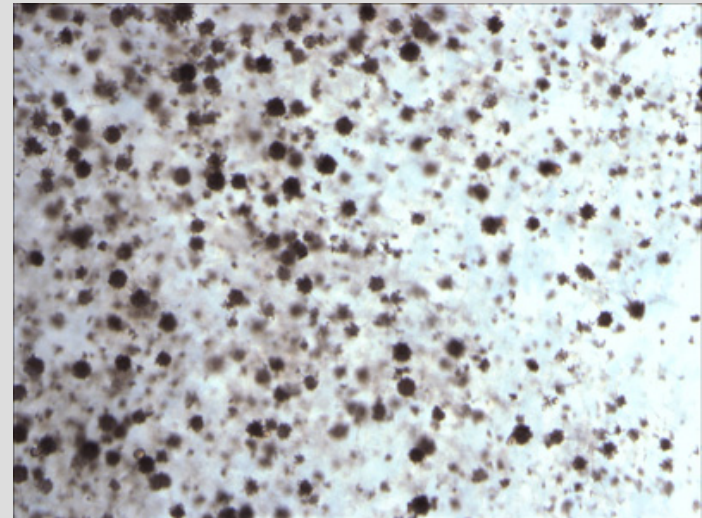
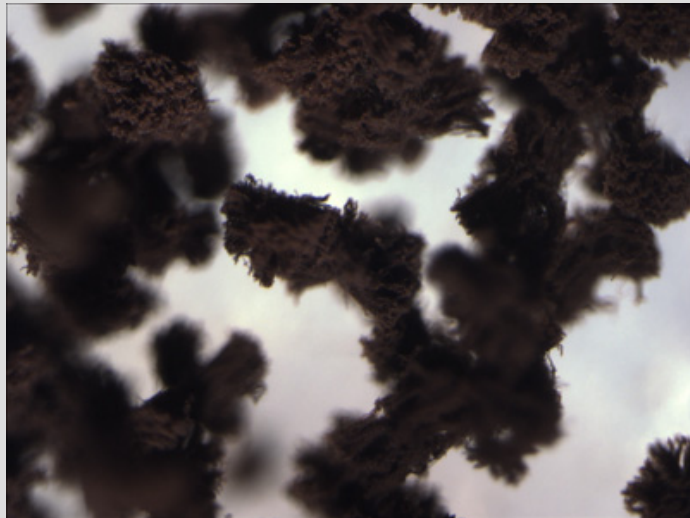
Wild-type (ATCC 9029)

NRRL 3122

~10X



~80X



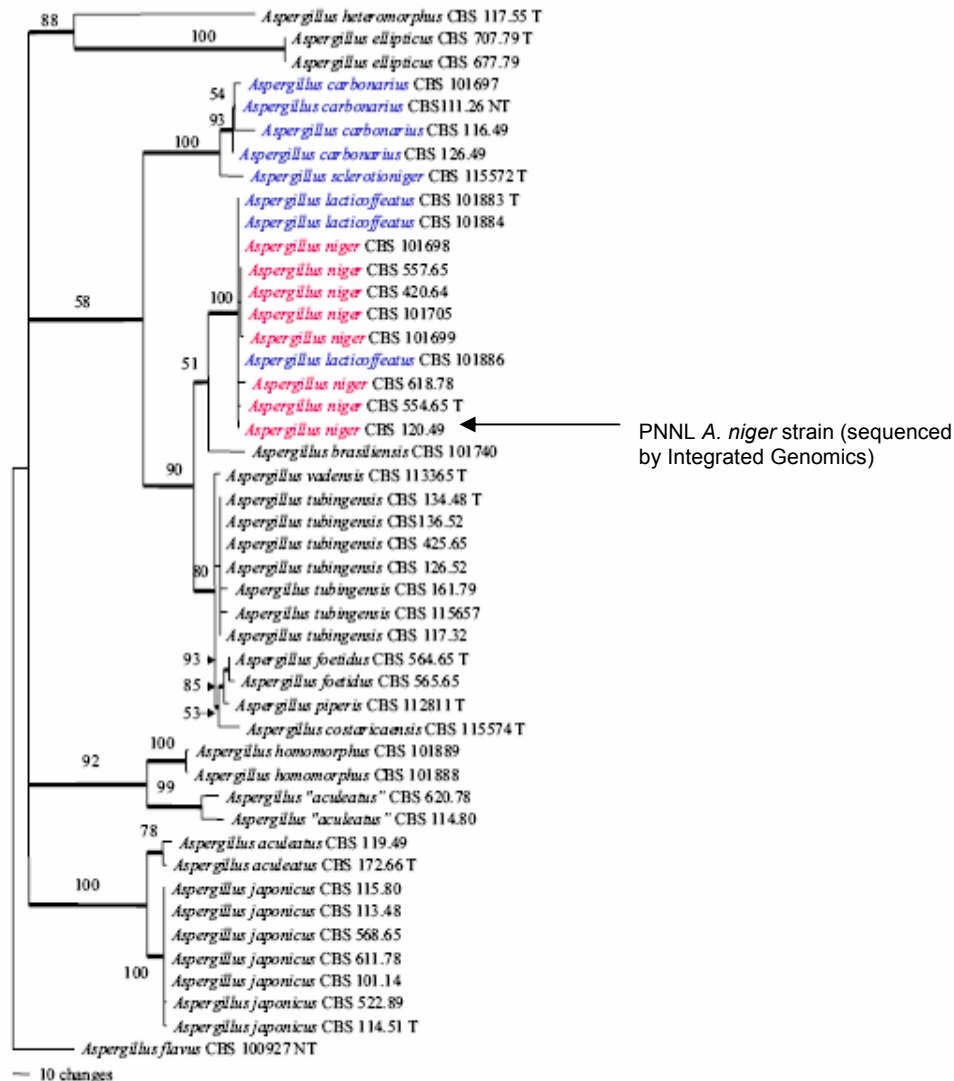


Fig. 1. One of the 5000 equally MPT of 719 steps based on heuristic search partial β -tubulin sequences with *A. flavus* as an outgroup. The branches in bold are 100 % in the 70 % majority-rule consensus of equally parsimonious trees. The numbers represent bootstrap percentages > 50 % (CI = 0.701, RI = 0.898 RC = 0.630, HI = 0.299). Names in blue are ochratoxin producing taxa. Taxa in red contain isolates which can produce ochratoxin.

Phylogeny from Robert A. Samson, Jos A.M.P. Houbaken, Angelina F.A. Kuijpers, J. Mick Frank and Jens C. Frisvad. 2004. New ochratoxin A or sclerotium producing species in *Aspergillus* section Nigri. Studies in Mycology. 50:45-61.

The DOE *Aspergillus niger* genome project

- ▶ Proposed to the US Department of Energy Microbial Genome Program by the PNNL Fungal Biotechnology team
- ▶ Collaboration with DOE's Joint Genome Institute
- ▶ Current status
 - Final draft coverage: ~8X shotgun
 - EST libraries constructed from RNA isolated from citric acid production and complex biomass digestion conditions ~30,000 sequenced
 - Annotation: In collaboration with JGI/LANL
 - Public release: Soon
- ▶ Other *Aspergillus niger* genomes
 - ATCC 9029: low coverage, sequenced by Integrated Genomics – Sequence available on request. Contact Scott Baker or Jon Magnuson (scott.baker@pnl.gov or jon.magnuson@pnl.gov)
 - CBS 513.88 (derivative of NRRL 3122/ATCC 22343): DSM – announced public release at Asilomar FGC

	Integrated Genomics	JGI
•Date	2000	2005
•Strain	Wild-type (ATCC 9029)	Wild-type (ATCC 1015)
•Method	Shotgun No finishing	Shotgun Plus finishing
•Coverage	~4-6X	~8X
•Genomic library insert size	1-2kb	3kb 8kb 40kb
•Contigs or scaffolds	>9000 contigs	<100 scaffolds

The “QC” *A. niger* ATCC 1015 assembly – 4X coverage

- ▶ total number of scaffolds: 118
- ▶ total length of scaffolds: 35634017
- ▶ N50 scaffold number: 6
- ▶ N50 scaffold size: 1931570
- ▶ total number of contigs: 1646
- ▶ total length of contigs: 34162656
- ▶ N50 contig number: 215
- ▶ N50 contig size: 47636
- ▶ Total: 243,688 reads
 - 3 kb: 105,065 = 43.1%
 - 8 kb: 118,655 = 48.7%
 - 40 kb: 19,968 = 8.2%

The “draft” *A. niger* ATCC 1015 assembly – 8X coverage

- ▶ Over 40 that encode ketosynthase and acyl-transferase domains(i.e. PKSs and FASs)
- ▶ Mat-1-1(alpha box)
- ▶ ~95% of the of the genome is found in 24 scaffolds – 1.5 scaffolds/chromosome arm
- ▶ EST coverage/annotation
 - Genencor to release ~7500 EST sequences from several different growth condition libraries
 - JGI sequenced 30,000 ESTs
 - The Fungal Genomics program at Concordia University will contribute ~12,000 ESTs
- ▶ Annotation jamboree scheduled for April 2006, following the European Conference on Fungal Genetics
- ▶ Limited gap closure or “finishing” is planned by JGI-LANL

What's next? A multi-gene phylogeny and more genomes from *Aspergillus* section *Nigri*

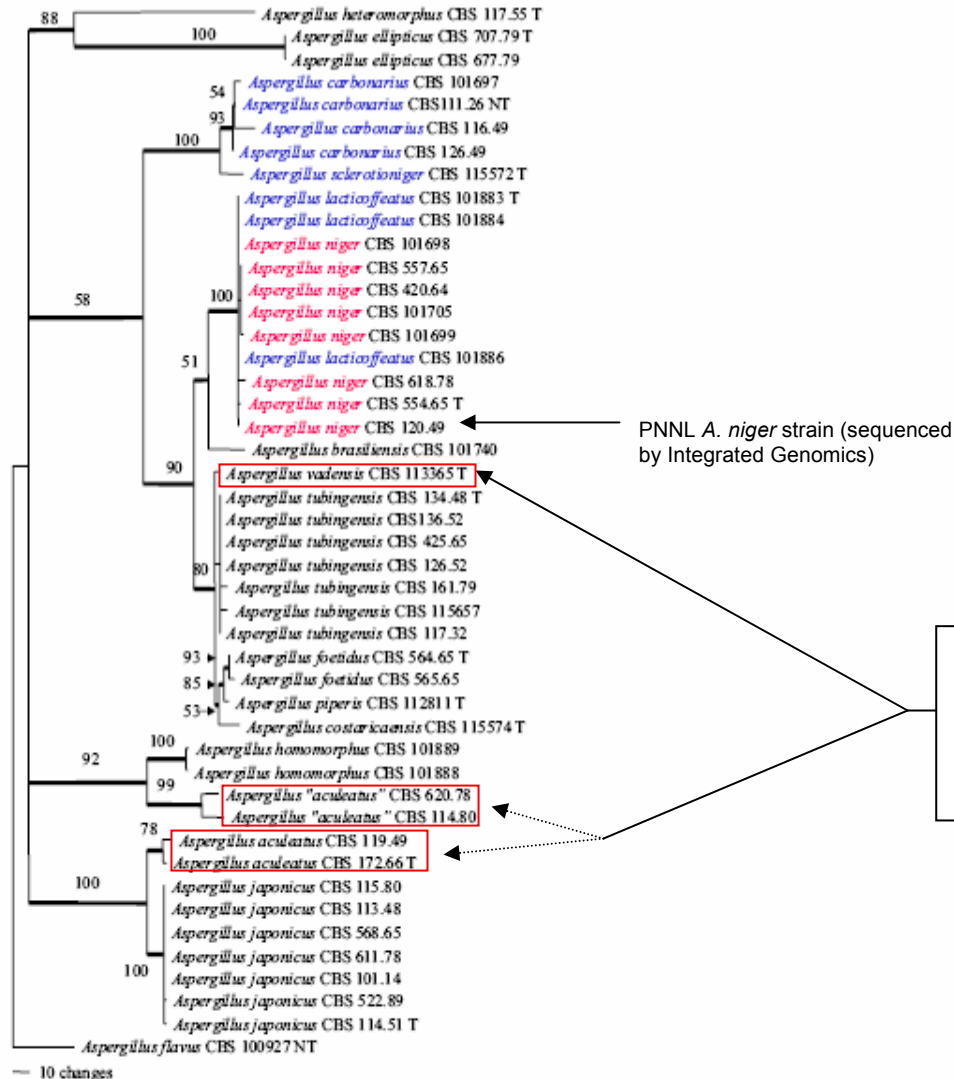
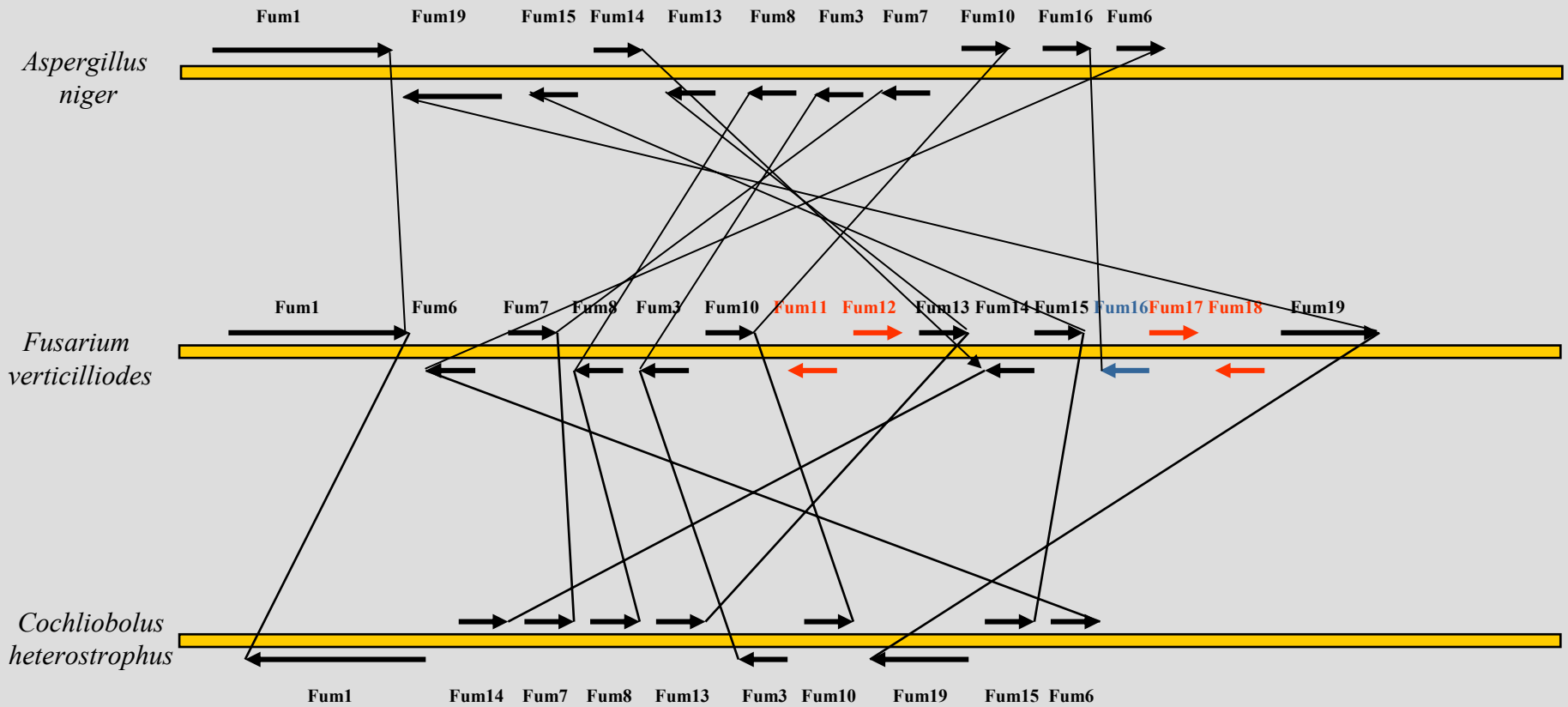


Fig. 1. One of the 5000 equally MPT of 719 steps based on heuristic search partial β -tubulin sequences with *A. flavus* as an outgroup. The branches in bold are 100 % in the 70 % majority-rule consensus of equally parsimonious trees. The numbers represent bootstrap percentages > 50 % (CI = 0.701, RI = 0.898 RC = 0.630, HI = 0.299). Names in blue are ochratoxin producing taxa. Taxa in red contain isolates which can produce ochratoxin.

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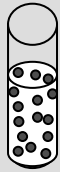
Conserved fumonisin-like gene clusters



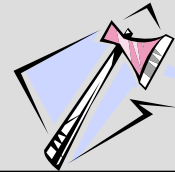
Future directions

- ▶ Cell biological studies of wild-type vs. “hyper producers”
- ▶ Comparative genomics of wild-type vs. “hyper producers”
- ▶ Phylogenetic studies of *Aspergillus niger* and relatives
- ▶ Medium throughput deletion of putative *A.niger* morphology genes

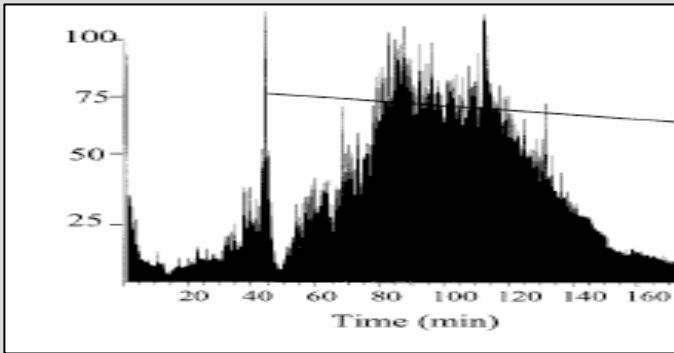
General Procedure for Proteomics



Lyse cells
Isolate proteins



Digest with trypsin

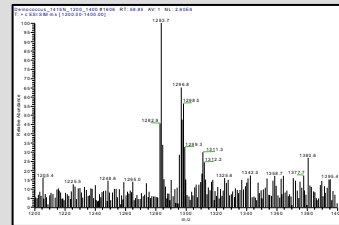


Separate in one or more dimensions
reverse phase, ion exchange

- Perform
- LCQ
- analysis

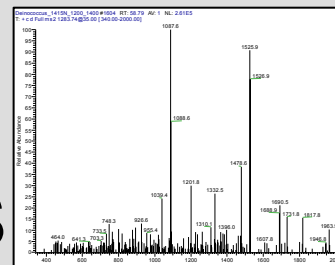
Raw Data

MS



Run data through peptide identifying program (SEQUEST)

Identify unique peptide = identify parent ORF



Quantitative proteomics

- ▶ Used for comparison of biological samples generated by two or more different experimental conditions
- ▶ Current technologies utilize isotopic labeling strategies
 - ICAT
 - Metabolic labeling
 - Pairwise comparison
- ▶ Our goal: Generate a quantitative proteomic methodology using statistical analysis of raw MS abundance data and that does not use isotopic labeling

Acknowledgements

PNNL Fungal Biotechnology Team

Ellen Panisko
Ken Bruno
Jon Magnuson
Ziyu Dai
Chris Wend
Beth Hofstad
Mark Butcher
Andy Zwoster
Debbie Lee
Kyle Fowler
Kelly Vincent
Katie Panther
Dennis Stiles
Linda Lasure

PNNL Proteomics QC Analysis Team

Don Daly
Kevin Anderson
Matt Monroe

DOE JGI *A. niger* ATCC 1015 genome

Dan Drell
Diego Martinez
Dan Rokhsar
Chris Detter
David Bruce
Erika Lindquist
Paul Richardson
...the list continues to grow!